

## WEST Search History





DATE: Sunday, March 14, 2004

Hide?	Set Name	Query	Hit Count
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*DB=USPT; PLUR=YES; OP=ADJ*

<input type="checkbox"/>	L19	l16 and L18	45
<input type="checkbox"/>	L18	l3 same l2	1191
<input type="checkbox"/>	L17	L16 and l1	56
<input type="checkbox"/>	L16	L15 same l2	67
<input type="checkbox"/>	L15	m7 or "m 7"	9198
<input type="checkbox"/>	L14	L13 and l1 and l6	33
<input type="checkbox"/>	L13	amino or nh2	240979
<input type="checkbox"/>	L12	(L11 same l2) and l1 and l6	12
<input type="checkbox"/>	L11	tenebrionis	274
<input type="checkbox"/>	L10	L9 and l6	15
<input type="checkbox"/>	L9	l2 and l3 and l1 not l7	343
<input type="checkbox"/>	L8	l2 and l7 and l1	16
<input type="checkbox"/>	L7	l5 and L6	16
<input type="checkbox"/>	L6	198?.pray.	355700
<input type="checkbox"/>	L5	L4 and l1	175
<input type="checkbox"/>	L4	L3 with l2	700
<input type="checkbox"/>	L3	truncat\$6 or delet\$5	157114
<input type="checkbox"/>	L2	toxin or endotoxin	24921
<input type="checkbox"/>	L1	thuringiensis.ti,ab,clm.	651

END OF SEARCH HISTORY



FILE 'REGISTRY' ENTERED AT 08:42:21 ON 14 MAR 2004  
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.  
PLEASE SEE "HELP USAGETERMS" FOR DETAILS.  
COPYRIGHT (C) 2004 American Chemical Society (ACS)  
STRUCTURE FILE UPDATES: 12 MAR 2004 HIGHEST RN 662722-88-5  
DICTIONARY FILE UPDATES: 12 MAR 2004 HIGHEST RN 662722-88-5

TSCA INFORMATION NOW CURRENT THROUGH JANUARY 6, 2004

FILE 'REGISTRY' ENTERED AT 08:42:21 ON 14 MAR 2004  
L1 54 S GWGPFGGALYS/QSP

FILE 'CA' ENTERED AT 08:42:58 ON 14 MAR 2004  
FILE COVERS 1907 - 11 Mar 2004 VOL 140 ISS 12  
FILE LAST UPDATED: 11 Mar 2004 (20040311/ED)

L2 19 S L1

FILE 'REGISTRY' ENTERED AT 08:43:35 ON 14 MAR 2004  
L3 49 S L1 AND 585-660/SQL

FILE 'CA' ENTERED AT 08:44:43 ON 14 MAR 2004  
L4 17 S L3

L2 ANSWER 1 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Identifying and reducing the allergenicity of food proteins  
PY 2003

L2 ANSWER 2 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Modified Cry3A toxins having increased toxicity to corn rootworm, their nucleic acid sequences, and methods for controlling plant pests  
PY 2003 2003 2003

L2 ANSWER 3 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Kits for the detection of transgenes in food plants for detection of genetically modified organisms in foodstuffs  
PY 2001 2002 2002

L2 ANSWER 4 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Fire ant control using a novel <SYM100>-endotoxin from *Bacillus thuringiensis*  
PY 2001 2002 2003 2002 2003

L2 ANSWER 5 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Substitution analogs of CryIIA <SYM100>-endotoxins with increased effectiveness against *Diabrotica*  
PY 1997

L2 ANSWER 6 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Synthetic insecticidal crystal protein gene for expression in transgenic plants  
PY 1996 2002 1999 2002 1995 1995 1996 1996 2000 2000 2004

L2 ANSWER 7 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Elucidation of the mechanism of CryIIA overproduction in a mutagenized strain of *Bacillus thuringiensis* var. *tenebrionis*  
PY 1994

L2 ANSWER 8 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 *Bacillus thuringiensis* isolates active against lice.  
PY 1993 1993 1993 1996 1994 1998 1998 1998

L2 ANSWER 9 OF 19 CA COPYRIGHT 2004 ACS on STN

FILE 'REGISTRY' ENTERED AT 08:47:05 ON 14 MAR 2004  
L5 1 S 110463-24-6/RN  
SET NOTICE 1 DISPLAY  
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L6 34 S L1 AND 585-643/SQL  
FILE 'CA' ENTERED AT 08:49:01 ON 14 MAR 2004  
L7 11 S L6

FILE 'REGISTRY' ENTERED AT 08:50:10 ON 14 MAR 2004  
L8 1 S 115804-12-1/RN  
SET NOTICE 1 DISPLAY  
SET NOTICE LOGIN DISPLAY

FILE 'REGISTRY' ENTERED AT 08:51:33 ON 14 MAR 2004  
L9 1 S 128123-81-9/RN  
SET NOTICE 1 DISPLAY  
SET NOTICE LOGIN DISPLAY

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L10 1 S 124541-32-8/RN  
SET NOTICE 1 DISPLAY  
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FILE 'REGISTRY' ENTERED AT 08:52:25 ON 14 MAR 2004  
L11 1 S 123514-67-0/RN

T1 The reconstruction and expression of a *Bacillus thuringiensis* cryIIA gene in protoplasts and potato plants  
PY 1993

L2 ANSWER 10 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Extending the host range of insecticidal proteins using peptides that bind gut cells  
PY 1991 1992 1991 1999 1992 1994

L2 ANSWER 11 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Synthetic insecticidal crystal protein gene  
PY 1990 1990 1995 1990 1995 1996 1996 1992 1990 2000 1990 1999 2002 2000 2000

L2 ANSWER 12 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Cloning and expression in microorganisms of endotoxin gene of *Bacillus thuringiensis* *tenebrionis*  
PY 1989 1989 1989 1989 1992 1990

L2 ANSWER 13 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Isolation and characterization of EG2158, a new strain of *Bacillus thuringiensis* toxic to coleopteran larvae, and nucleotide sequences of the toxin gene  
PY 1988

L2 ANSWER 14 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Plants transformed with a gene for an insecticidal protein from *Bacillus thuringiensis*  
PY 1989 1989 1989 1989 1989 1990

L2 ANSWER 15 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Cloning of *Bacillus thuringiensis* *tenebrionis* toxin gene and its use in producing coleopteran insect-resistant plants  
PY 1988 1989 1996 2003 1996 1996 1997 1988 1988 1991 1988 1994 1988 1998 1989 1994 2001 1996 1998 2001 1998 2001 2002

L2 ANSWER 16 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Characterization of the coleopteran-specific protein gene of *Bacillus thuringiensis* var. *tenebrionis*  
PY 1988

SET NOTICE 1 DISPLAY  
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FILE 'REGISTRY' ENTERED AT 08:53:08 ON 14 MAR 2004  
L12 1 S 123516-40-5/RN  
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SET NOTICE LOGIN DISPLAY

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L13 1 S 120313-97-5/RN  
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FILE 'REGISTRY' ENTERED AT 08:54:15 ON 14 MAR 2004

FILE 'REGISTRY' ENTERED AT 08:58:09 ON 14 MAR 2004  
L14 1 S 112659-37-7/RN  
SET NOTICE 1 DISPLAY  
SET NOTICE LOGIN DISPLAY

FILE 'REGISTRY' ENTERED AT 08:58:50 ON 14 MAR 2004  
L15 1 S 112659-38-8/RN  
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L2 ANSWER 17 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Molecular cloning and characterization of the insecticidal crystal protein gene of *Bacillus thuringiensis* var. *tenebrionis*  
PY 1987

L2 ANSWER 18 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Nucleotide sequence of a gene encoding an insecticidal protein of *Bacillus thuringiensis* v *tenebrionis* toxic against *Coleoptera*  
PY 1987

L2 ANSWER 19 OF 19 CA COPYRIGHT 2004 ACS on STN  
T1 Cloning and expression of *Bacillus thuringiensis* toxin gene toxic to beetles of the order *Coleoptera*  
PY 1987 1991 1989 1988 1987 1996 1990 1989 1991 1991 1996 1999 1996 1999 1995 1999

L4 ANSWER 1 OF 17 CA COPYRIGHT 2004 ACS on STN  
T1 Identifying and reducing the allergenicity of food proteins  
PY 2003

L4 ANSWER 2 OF 17 CA COPYRIGHT 2004 ACS on STN  
T1 Modified Cry3A toxins having increased toxicity to corn rootworm, their nucleic acid sequences, and methods for controlling plant pests  
PY 2003 2003 2003

L4 ANSWER 3 OF 17 CA COPYRIGHT 2004 ACS on STN  
T1 Fire ant control using a novel <SYM100>-endotoxin from *Bacillus thuringiensis*  
PY 2001 2002 2003 2002 2003

L4 ANSWER 4 OF 17 CA COPYRIGHT 2004 ACS on STN  
T1 Substitution analogs of CryIIA <SYM100>-endotoxins with increased effectiveness against *Diabrotica* PY 1997

L4 ANSWER 5 OF 17 CA COPYRIGHT 2004 ACS on STN  
T1 Synthetic insecticidal crystal protein gene for expression in transgenic plants  
PY 1996 2002 1999 2002 1995 1995 1996 1996 2000 2000 2004

L4 ANSWER 6 OF 17 CA COPYRIGHT 2004 ACS on STN



TI Elucidation of the mechanism of CryIIIA overproduction in a mutagenized strain of *Bacillus thuringiensis* var. tenebrionis  
PY 1994

L4 ANSWER 7 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI *Bacillus thuringiensis* isolates active against lice  
PY 1993 1993 1993 1996 1998 1998  
RN 110463-24-6 RN 130067-79-7 RN 134944-24-4 RN 141033-45-0 RN 141467-09-6  
RN 151404-48-7 RN 151404-50-1 RN 151404-53-4 RN 134945-86-1 RN 141467-29-0  
RN 151404-45-4 RN 151404-46-5 RN 151404-47-6 RN 151404-49-8 RN 151404-51-2  
RN 151404-52-3

L4 ANSWER 8 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI The reconstruction and expression of a *Bacillus thuringiensis* cryIIIA gene in protoplasts and potato plants  
PY 1993

L4 ANSWER 9 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Synthetic insecticidal crystal protein gene  
PY 1990 1990 1995 1990 1995 1996 1996 1990 1992 1990 2000  
RN 115804-12-1 RN 128123-81-9 RN 128123-03-5 RN 128123-04-6

L4 ANSWER 10 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Cloning and expression in microorganisms of endotoxin gene of *Bacillus thuringiensis* tenebrionis  
PY 1993

L5 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN  
RN 110463-24-6 REGISTRY  
CN Toxin, (*Bacillus thuringiensis* strain M-7 clone pCH-B3 parasporal crystal precursor reduced) (901) (CA INDEX NAME)  
OTHER NAMES:  
CN <SYM100>-Endotoxin CryIIIA (*Bacillus thuringiensis* tenebrionis strain NB176)  
CN Toxin, <SYM100>-endo- (*Bacillus thuringiensis* tenebrionis strain NB176 gene cryIIIA parasporal crystal)  
FS PROTEIN SEQUENCE  
SQL 644

SEQ 1 MNPNRSEHD TIKTENNEV PTNHVQYPLA ETPNPTLED NYKEFLRMTA  
51 DNNTALDSS TTKDVQKGI SWGDLGW GFPPGALVS FYTNFTIWI  
101 PSEDPWKAFM EQEALMDOK IADYAKNKAL AELOGLQNV EDYVSALSSW  
151 QKNPVSRNP HSQGRIRLF SOAESHFNRNS MPFSAISGYE VLFLTTYQA  
201 ANTHFLFKD AIGYEEWYG EKEDIAFYK RQLKLTQET DHCKVKNVG  
251 LDKLRGSSYE SWYFNRYRR EMTLTVDLI ALFPLYOYRL YPKEVKTSL  
301 RDVLTDPVG VNNLRGYGT FSNENYRK PHLFDYLRH QFHTRFQPGY  
351 YGNDSFNYS GNYVSTRPSI GSNDIITSPF YGNKSSEPVQ NLEFNGEKVY  
401 RAVANTNLAV WPSAVYSGVT KVEFSQNDQ IDEASTQYD SKRNVGAYSW  
451 DSIDLQPPET DDEPLEKGYH HQLNYVMCFL MQGSRGTIPV LVTWHSVDF  
501 FNMIDSKIT QLPVKAYKL QSGASWAGP RFTGGDIQC TENGSAATY  
551 VTPDVSYSQK YRARIHYAST SQITFTSLD GAPFNQYFD KTNKGDTLT  
601 YNSFNLASFS TPFELSGNNL QIGVTGLSAG DKVYIDKIEP IPVN

\*\*RELATED SEQUENCES AVAILABLE WITH SEQLINK\*\*  
MF Unspecified  
CI MAN  
SR CA  
LC STN Files: CA, CAPLUS, TOXCENTER, USPATFULL  
7 REFERENCES IN FILE CA (1907 TO DATE)  
1 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA  
7 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L7 ANSWER 1 OF 11 CA COPYRIGHT 2004 ACS on STN  
TI Modified Cry3A toxins having increased toxicity to corn rootworm, their nucleic acid sequences, and methods for controlling plant pests  
PY 2003 2003 2003

PI 1989 1989 1989 1992 1990  
RN 110463-24-6 RN 124541-32-8 RN 112659-24-2 RN 124541-05-5 RN 124541-04-4

L4 ANSWER 11 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Isolation and characterization of EG2158, a new strain of *Bacillus thuringiensis* toxic to coleopteran larvae, and nucleotide sequence of the toxin gene  
PY 1988  
RN 110463-24-6 RN 123514-67-0 RN 112659-24-2

L4 ANSWER 12 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Plants transformed with a gene for an insecticidal protein from *Bacillus thuringiensis*  
PY 1989 1989 1989 1989 1989 1989  
RN 123516-40-5 RN 123516-11-0 RN 62213-36-9

L4 ANSWER 13 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Cloning of *Bacillus thuringiensis* tenebrionis toxin gene and its use in producing coleopteran insect-resistant plants  
PY 1988 1988 1996 2003 1996 1997 1988 1988 1991 1988  
RN 1994 1988 1988 1989 1994 2001 1996 1998 2001 1998 2001

RN 120313-96-4 RN 120313-97-5 RN 120313-98-6 RN 120313-99-7 RN 120314-00-3  
RN 120314-01-4 RN 9027-23-0 RN 108281-08-9 RN 120313-68-0 RN 120313-69-1  
RN 120313-70-4

L4 ANSWER 14 OF 17 CA COPYRIGHT 2004 ACS on STN

L7 ANSWER 2 OF 11 CA COPYRIGHT 2004 ACS on STN  
TI Fire ant control using a novel <SYM100>-endotoxin from *Bacillus thuringiensis*  
PY 2001 2002 2003 2002 2003

L7 ANSWER 3 OF 11 CA COPYRIGHT 2004 ACS on STN  
TI Synthetic insecticidal crystal protein gene for expression in transgenic plants  
PY 1996 2002 1999 2002 1995 1996 2000 2000 2000 2004

L7 ANSWER 4 OF 11 CA COPYRIGHT 2004 ACS on STN  
TI The reconstruction and expression of a *Bacillus thuringiensis* cryIIIA gene in protoplasts and potato plants  
PY 1993

L7 ANSWER 5 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 113-38937 CA  
TI Synthetic insecticidal crystal protein gene  
IN Adang, Michael J.; Rocheleau, Thomas A.; Merlo, Donald J.; Murray, Elizabeth E.  
PA Lubrizol Genetics, Inc., USA  
SO Eur. Pat. Appl., 30 pp. CODEN: EPXXDW  
DT Patent LA English  
FAN ONT 7

PATENT NO. KIND DATE APPLICATION NO. DATE

PI EP 359472 A2 19900321 EP 1989-309069 19890907  
EP 359472 A3 19900725  
EP 359472 B1 19951227  
R: AT, BE, CH, DE, ES, FR, GB, GR, IT, LI, LU, NL, SE  
ZA 8906562 A 19900530 ZA 1989-6562 19890828  
EP 682115 A1 19951115 EP 1995-201374 19890907  
R: AT, BE, CH, DE, ES, FR, GB, GR, IT, LI, LU, NL, SE  
AT 132193 E 19960115 AT 1989-309069 19890907  
ES 2083384 T3 19960416 ES 1989-309069 19890907  
AU 8941182 A1 19900315 AU 1989-41182 19890908  
AU 623429 B2 19920514  
CN 1044298 A 19900801  
CN 1056880 B 20000927  
JP 02186989 A2 19900723  
JP 11266882 A2 19991005  
JP 2002176995 A2 20020625  
US 6013523 A 20000111  
CN 1263946 A 20000823  
PRAI US 1988-242482 A 19880909

embio U.S. priority 9/86

TI Characterization of the coleopteran-specific protein gene of *Bacillus thuringiensis* var. tenebrionis  
PY 1988  
RN 115804-12-1 RN 112659-37-7 RN 112659-24-2 RN 115803-70-8

L4 ANSWER 15 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Molecular cloning and characterization of the insecticidal crystal protein gene of *Bacillus thuringiensis* var. tenebrionis  
PY 1987  
RN 110463-24-6 RN 112659-24-2

L4 ANSWER 16 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Nucleotide sequence of a gene encoding an insecticidal protein of *Bacillus thuringiensis* var. tenebrionis toxic against Coleoptera  
PY 1987  
RN 112659-37-7 RN 112659-38-8 RN 112659-24-2

L4 ANSWER 17 OF 17 CA COPYRIGHT 2004 ACS on STN  
TI Cloning and expression of *Bacillus thuringiensis* toxin gene toxic to beetles of the order Coleoptera  
PY 1987 1991 1989 1988 1987 1996 1990 1989 1991 1991  
RN 1996 1999 1996 1999 1996 1999

RN 110463-24-6



EP 1989-309069 A3 19890907  
JP 1989-235472 A3 19890911  
JP 1988-356822 A3 19890911  
US 1992-827844 B1 19920128  
US 1993-57191 A3 19930503  
US 1995-369839 A3 19950106  
RN 115804-12-1 RN 128123-81-9 RN 128123-03-5 RN 128123-04-5

*U8*  
*SG1*  
*SG1*

L7 ANSWER 6 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 11231703 CA  
TI Cloning and expression in microorganisms of endotoxin gene of *Bacillus thuringiensis tenebrionis*  
IN Sekar, Vaitilingham; Adang, Michael J.  
PA Lubrizol Genetics, Inc., USA  
SO Eur. Pat. Appl. 29 pp. CODEN: EPXXDW  
DT Patent LA English  
FAN CNT 1

PATENT NO.	KIND DATE	APPLICATION NO.	DATE
PI EP 318143	A2 19890531	EP 1988-309438	19881010
EP 318143	A3 19890607		
ZA 8807480	A 19890726	ZA 1988-7480	19881005
AU 8823651	A1 19890413	AU 1988-23651	19881012
AU 626804	B2 19920813		
JP 02092287	A2 19900403	JP 1988-259289	19881013
PRAI US 1987-108285	19871013		
RN 110463-24-6 RN 124541-32-6 RN 112659-24-2 RN 124541-05-5 RN 124541-04-4			

*U8*  
*SG1*  
*SG1*

L7 ANSWER 7 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 111209800 CA  
TI Isolation and characterization of EG2158, a new strain of *Bacillus thuringiensis* toxic to coleopteran larvae, and nucleotide sequence of the toxin gene  
AU Donovan, William P.; Gonzalez, Jose M.; Gilbert, M. Pearce; Dankocsik, Cathy  
CS Ecogen Inc., Langhorne, PA, 19047, USA  
SO Molecular and General Genetics (1988), 214(3), 365-72 CODEN: MGGEAE; ISSN: 0026-8925  
DT Journal LA English  
RN 110463-24-6 RN 123514-67-0 RN 112659-24-2

L7 ANSWER 8 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 111188783 CA  
TI Plants transformed with a gene for an insecticidal protein from *Bacillus thuringiensis*  
IN Vaeck, Mark; Hofte, Hermanus; Botterman, Johan  
PA Plant Genetic Systems N. V., Belg.  
SO Eur. Pat. Appl. 22 pp. CODEN: EPXXDW  
DT Patent LA English  
FAN CNT 1

PATENT NO.	KIND DATE	APPLICATION NO.	DATE
PI EP 305275	A2 19890301	EP 1988-402115	19880816
EP 305275	A3 19890614		
R: AT, BE, CH, DE, ES, FR, GB, GR, IT, LI, LU, NL, SE			
WO 8901515	A2 19890223	WO 1988-EP752	19880815
WO 8901515	A3 19890601		
W: AU, JP, US			
AU 8822572	A1 19890309	AU 1988-22572	19880815
JP 02500566	T2 19900301	JP 1988-506676	19880815
PRAI GB 1987-19414	19870817		
GB 1987-30261	19871229		
WO 1988-EP752	19880815		
RN 123516-40-5 RN 123516-11-0 RN 62213-36-9			

L7 ANSWER 9 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 110187338 CA

TI Cloning of *Bacillus thuringiensis tenebrionis* toxin gene and its use in producing coleopteran insect-resistant plants  
IN Fischhoff, David Allen; Fuchs, Roy Lee; McPherson, Sylvia Ann; Lavrik, Paul Bruno; Perlak, Frederick Joseph  
PA Monsanto Co., USA  
SO Eur. Pat. Appl. 52 pp. CODEN: EPXXDW  
DT Patent LA English  
FAN CNT 1

PATENT NO.	KIND DATE	APPLICATION NO.	DATE
PI EP 289479	A2 19881102	EP 1988-870070	19880426
EP 289479	A3 19890628		
EP 289479	B1 19961106		
EP 289479	B2 20030910		
R: AT, BE, CH, DE, ES, FR, GB, GR, IT, LI, LU, NL, SE			
EP 731170	A1 19960911	EP 1996-100978	19880426
AT 144995	E 19961115	AT 1988-870070	19880426
ES 2094722	T3 19970201	ES 1988-870070	19880426
DK 8602340	A 19881031	DK 1988-2340	19880428
AU 8815273	A1 19881103	AU 1988-15273	19880428
AU 610157	B2 19910516		
CN 88102497	A 19881123	CN 1988-102497	19880428
CN 1026497	B 19941109		
JP 63287488	A2 19881124	JP 1988-107503	19880428
JP 2815047	B2 19981027		
ZA 8803049	A 19890222	ZA 1988-3049	19880428
RU 2025486	C1 19941230	RU 1988-435607	19880428
JP 2001112490	A2 20010424	JP 2000-272128	19880428
US 5495071	A 19960227	US 1993-72281	19930604
US 5763241	A 19980609	US 1996-759446	19961205
US 6284949	B1 20010904	US 1998-27998	19980223
JP 10323138	A2 19981208	JP 1998-101210	19980413
JP 3122642	B2 20010109		
US 2002152496	A1 20021017	US 2001-943692	20010831
PRAI US 1987-44081	A 19870429		
EP 1988-870070	A3 19880426		
JP 1988-107503	A3 19880428		
JP 1998-101210	A3 19880428		
US 1990-523284	B1 19900514		
US 1993-72281	A3 19930604		
US 1995-435101	B1 19950504		
US 1996-759446	A1 19961205		
US 1998-27998	A3 19980223		
RN 120313-96-4 RN 120313-97-5 RN 120313-98-6 RN 120313-99-7 RN 120314-00-3 RN 120314-01-4 RN 9027-23-0			
RN 108281-08-9 RN 120313-68-0 RN 120313-69-1 RN 120313-70-4			

L7 ANSWER 10 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 10987114 CA  
TI Characterization of the coleopteran-specific protein gene of *Bacillus thuringiensis* var. *tenebrionis*  
AU McPherson, Sylvia A.; Perlak, Frederick J.; Fuchs, Roy L.; Marrone, Pamela G.; Lavrik, Paul B.; Fischhoff, David A.  
CS Monsanto Co., St. Louis, MO, 63198, USA  
SO BiorTechnology (1988), 6(1), 61-6 CODEN: BTCHDA; ISSN: 0733-222X  
DT Journal LA English  
RN 115804-12-1 RN 112659-37-7 RN 112659-24-2 RN 115803-70-8

L7 ANSWER 11 OF 11 CA COPYRIGHT 2004 ACS on STN  
AN 10869763 CA  
TI Nucleotide sequence of a gene encoding an insecticidal protein of *Bacillus thuringiensis* var. *tenebrionis* toxic against Coleoptera  
AU Hoeft, Herman; Seurinck, Jef; Van Houtven, Annemie; Vaeck, Mark  
CS Plant Genet. Syst., N. V., Ghent, 9000, Belg.  
SO Nucleic Acids Research (1987), 15(17), 7183 CODEN: NARHAD; ISSN: 0305-1048  
DT Journal LA English  
RN 112659-37-7 RN 112659-38-8 RN 112659-24-2

*pub 9/87*



L8 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 115804-12-1 REGISTRY

CN Toxin, endo- (Bacillus thuringiensis tenebrionis plasmid clone pMON5456 parasporal crystal reduced) (9Ci)

(CA INDEX NAME)

FS PROTEIN SEQUENCE

SQL 597

SEQ 1 MTADNNTAAL DSSTTKDVIQ KGISVVGDL GVVGFPPGGA LVSFYTNFLN  
51 TIWPSDPWK AFMEQVEALM DQKIADYAKN KALAEQLGLQ NNVEDYVSA  
101 SSWQKNPVSS RNPHSQGRIR ELFSQAESHF RNSMPFSAIS GYEVFLTTY  
151 AQAANTNLF LKDAQYIGE WGYEKEDIAE FYKRQLKLTQ EYTDHCVKWY  
201 NVGLDKLRGS SYESWVNWFR YRREMTLTVL DLIALFLYD VRLYPKEVKT  
251 ELTRDVLTD IPGVNNLRGY GTTFSENIEN IRKPHLFDYL LHRIQFHTRF  
301 PGYYGNDNFSN YWGSNYSVSTR PSIGSDIIT SPFYGNKSS EPVQNLFEFG  
351 KYVRAVANTN LAWPSAVYS GVTKEVFSQY NDQIDEASTQ TYDSKRNVGA  
401 VSWDSIDLQ PETTDEPLEK GYSHQLNVM CFLWQGSRGTP IPIVLTWTHKS  
451 VDFENMIDSK KITQLPLVKA YKLQSGASV AGPRFTGGDI IQCTENGSA  
501 TIYVTPDVS YSQYRARIH ASTSQITFTL SLDGAPFNQY YFDKTINKGD  
551 TLTYSNFNLA SFSTPFELSG NNLIQVGTGL SAGDKVYIDK IEFIPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

2 REFERENCES IN FILE CA (1907 TO DATE)

2 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L9 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 128123-81-9 REGISTRY

CN Toxin, endo- (Bacillus thuringiensis tenebrionis plasmid clone pMON5456 parasporal crystal reduced) 2-L-

alanine-597a-L-leucine-597b-L-arginine-597c-L-serine-597d-L-proline-597e-glycine-597f-L-threonine-597g-L-

glutamic acid-597h-L-leucine-597i-L-glutamic acid-597j-L-phenylalanine-597k-L- isoleucine-597l-aspartic acid-

597m-L-isoleucine- (9Ci) (CA INDEX NAME)

FS PROTEIN SEQUENCE

SQL 610

SEQ 1 HAADHWTEAL DSSTTKDVIQ KGISVVGDL GVVGFPPGGA LVSFYTWFLW  
51 TIWPSDPWK AFMEQVEALM DQKIADYAKW KALAEQLGLQ WHVEDYVSA  
101 SSWQKNPVSS RHPSQGRIR ELFSQAESHF RWSMPFSAIS GYEVFLTTY  
151 AQAANTNLF LKDAQYIGE WGYEKEDIAE FYKRQLKLTQ EYTDHCVKWY  
201 WGLDKLRGS SYESWVNWFR YRREMTLTVL DLIALFLYD VRLYPKEVKT  
251 ELTRDVLTD IPGVNNLRGY GTTFSENIEN YRKPPLFDY LNRIQFHTRF  
301 PGYYGNDNFSN YWGSNYSVSTR PSIGSDIIT SPFYGNKSS EPVQNLFEFG  
351 KYVRAVANTN LAWPSAVYS GVTKEVFSQ YMDQTDASTQ TYDSKRNVGA  
401 VSWDSIDLQ PETTDEPLEK GYSMLMYVM CFLWQGSRGTP IPIVLTWTHKS  
451 VDFENMIDSK KITQLPLVK AYKLQSGASV VAGPRFTGGDI IQCTENGSA  
501 TIYVTPDVS YSQYRARIH YASTSQITFTL SLDGAPFNQY YFDKTINKGD  
551 TLTYSNFMILA SFSTPFELSG WWLIQVGTGL SAGDKVYIDK IEFIPVHLRS  
601 PGTELEFIDI

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L10 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 124541-32-8 REGISTRY

CN 47-644-Toxin, endo- (Bacillus thuringiensis tenebrionis parasporal crystal precursor reduced) (9Ci) (CA

INDEX NAME)

FS PROTEIN SEQUENCE

SQL 598

SEQ 1 RMTADNNTAAL DSSTTKDVI QKGISVVGDL LGVGFPPGGA ALVSFYTNFL  
51 TIWPSDPWK AFMEQVEAL MDQKIADYAK KALAEQLGLQ QNNVEDYVSA  
101 SSWQKNPVSS RNPHSQGRIR ELFSQAESHF RNSMPFSAIS GYEVFLTTY  
151 YQAANTHLF LKDAQYIGE EWGYEKEDIA EFKRQLKLT QEYTDHCVKW  
201 YNVGLDKLRG SSYESWVWNFR YRREMTLTV DLIALFLYD VRLYPKEVKT  
251 TELTRDVLTD IPGVNNLRG YGTTFSENIEN YRKPPLFDY LHRIQFHTRF  
301 PGYYGNDNFSN YWGSNYSVSTR PSIGSDIIT SPFYGNKSS EPVQNLFEFG  
351 EKVYRAVANTN LAWPSAVYS GVTKEVFSQ YNDQIDEAST QTYDSKRNVG  
401 AVSWDSIDLQ PETTDEPLE KGYSHQLNVM MCELMQGSRG TIPVLTWTHK  
451 SVDFENMIDSK KITQLPLVK AYKLQSGASV VAGPRFTGGDI IQCTENGSA  
501 ATYVTPDVS YSQYRARIH YASTSQITFTL SLDGAPFNQY YFDKTINKG  
551 DLTYSNFNLA SFSTPFELSG GNNLIQVGTGL SAGDKVYIDK IEFIPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L11 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 123514-57-0 REGISTRY

CN Toxin, endo- (Bacillus thuringiensis strain EG2158 plasmid clone pEG212 parasporal crystal reduced) (9Ci)

(CA INDEX NAME)

FS PROTEIN SEQUENCE

SQL 594

SEQ 1 TEALDSSTTK DVIQKGISVW GDLLGVVGFP FGGALVSFYT NFLNTWPSE  
51 DPWKAFMEQV EALMDQKID YAKNKALAE QGLQNNVEDY VSALSSWQKN  
101 PVSSRNPHSQ GRIRSELFQA ESHFRNSMPS TAISGYEVLF LTTYAQAANT  
151 HLFLDKAQI YGEEWGYEKE DIAEFYKRQL KLTQETDHC VKWYNVGLDK  
201 LRSSYESWV NFNRYRREMT LTVLDIALF PLYDVRLYPK EVKTELTRDV  
251 LTPDIVGNN LREYGTTFSN IENYRKPPL FDLHRIQFH TRFQPGYYGN  
301 DSFNWWSGNY VSTRPSIGN DIITSPFYGN KSSGPVQNL ENGEKYVRAY  
351 ANTNLAWPS AVYSGVTKEV FSQYNDQIDE ASTQTYDSKR NVGASWDSI  
401 DQLPETTDE PLEKGYSHQL NYVMCFLMQG SRGTIPVLTW THKSVDFENM  
451 IDSKKITQLP LVKAYKAYKL QSGASVWAGP RFTGGDIQC TENGSAATY  
501 VTPDVSYSQK YRARIHYAST SQITFTLSLD GAPFNQYFDT KTINKGDTLT  
551 YNSFNLASFS TPFELSGNNL QIGVTGLSAG DKVYIDKIEF IPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)



L12 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN  
RN 123516-40-5 REGISTRY  
CN 8-594-Toxin, endo- (Bacillus thuringiensis tenebrionis parasporal crystal reduced (9CI)) (CA INDEX NAME)  
FS PROTEIN SEQUENCE  
SQL 587

SEQ 1 DSSTTKDVIQ KGISVVGDL GVVGFPPGGA LVSYFTNFLN TIWPSDPWK  
51 AFMEQVEALM DOKIADYAKN KALAELOGLQ NNVEDYVSAL SSWQKNPVSS  
101 RNPHSQGRIR ELFSQAESHF RNSMPSFAIS GYEVLFLLTY AQAANTHLFL  
151 LKDAQIYGEE WGYEKEDIAE FYKRQLKLTQ EYTNHCWKWY NVGLDKLRGS  
201 SYESWVNFNR YREMTLTVL DLALFLYD VRLYPKEVKT ELTRDVLTD  
251 IGVNVLRGY GTTFSNIENY IRKPHLFDYL HRIQFHTRFQ PGYYGNDSFN  
301 YWSGNYVSTR PSIGSNDIT SPFYGNKSSSE PIVQNEFNCE KYRAVANTN  
351 LAWPSAVYS GYTKVEFSQY NDQIDEASTQ TYDSKRNUGA VSWDSIDQLP  
401 PETTDEPLEK GYSHQLNYVM CFLMQGSRGT IPVLTVTHKS VDFNMDISK  
451 KITQLPLVKA YKLQSGASV AGPRFTGGDI IQCTENGSAATYVTPDVSY  
501 SQKYRARIHY ASTSQITFTL SLDGAPFNQY YFDKTINKGD TLTYNSFNL  
551 SFSTPFELSG NNLQIGVTGL SAGDKYVIDK IEFIPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L13 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 120313-97-5 REGISTRY

CN 16-644-Toxin, endo- (Bacillus thuringiensis tenebrionis plasmid clone pMON5420/pMON5421 parasporal crystal precursor reduced) (9CI) (CA INDEX NAME)

FS PROTEIN SEQUENCE

SQL 629

SEQ 1 ENNEVPTNHV QYPLAETPNP TLEDNLNKEF LRMTADNNTA ALDSSTTKDV  
51 IQKGISVVGD LLGVVGFPPG GALVSFYTNF LNTIWPSDDP WKAFMEQVEA  
101 LMDQKIADYA KKKALAELOQ LQNNVEDYVS ALSSWQKNPV SSRNPHSQGR  
151 IRELFQSAES HFRNSMPSFA ISGYEVFLT TYAQAANTHL FLLKDAQIYG  
201 EEWGYEKEDIAE FYKRQLKLTQ EYTNHCWKWY NVGLDKLR GSSYESWVNF  
251 NRYRREMTLT VLDLALFPL YDRLYPKEV KTELTRDVLTDPIVGVNLR  
301 GYGTTFESNIE NYIRKPHLFD YLHRIQFTR FQPGYYGNDN FNYWSGNYVS  
351 TRPSIGSNDI ITSPFYGNKS SEPQVQNEFN GEKVYRAVAN TNLAVWPSAV  
401 YSGVTKEFS QYNDQIDEAS TQTYDSKRN VAYSWDSIDQLP PETTDEPL  
451 EKGYSHELNY VMCFLMQGSR GTIPVLTVTH KSVDFENMD SKKITQLPLV  
501 KAYKLQSGAS WAGPRFTGG DIQCTENGSAATYVTPDV SYSQKYRARI  
551 HYASTSQITF TSLSDGAPFN QYYFDKTINK GDTLTYNSFN LASFSTPFEL  
601 SGNNLQIGVT GLSAGDKYVIDK IEFIPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER, USPATFULL

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L14 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN  
RN 112659-37-7 REGISTRY  
CN Toxin, endo- (Bacillus thuringiensis tenebrionis parasporal crystal precursor reduced) (9CI) (CA INDEX NAME)  
FS PROTEIN SEQUENCE  
SQL 643

SEQ 1 MNPNNRSEHD TIKTTENNEV PTNHVQYPLA ETPNPTLEDL NYKEFLRMTA  
51 DNNTAELDSS TTKDVIQKGI SVVGDLGGV GFPPGGALVS FYTNFLNTIW  
101 PSEDPWKAFM EQVEALMDQK IADYAKNKAL AELQGLQNNV EDYVSALSSW  
151 QKNPVSSRNP HSQGRIRELF SQAESHFRNS MPSFAISGYE VLFLTTYAQA  
201 ANTHLFLKLD AQYGEWGY EKEDIAEFYK RQLKLTQEYT DHCVKWYNVG  
251 LDKLRGSSYE SWVNFNRYRR EMTLTVLDLI ALFPLYDVRL YPKEVKTELT  
301 RDVLTDPIVG VNNLRGYGT FSNENYIRKP HLFDYLHRIQ FHTFRFQPGY  
351 GNDSFNWSG NYVSTRPSIG SNDIITSPFY GNKSSEPQVQ NEFNGEKVYR  
401 AVANTNLAVW PSAYSGVTK VEFQYNDQT DEASTQTYDS KRNVGAVSWD  
451 SIDQLPETT DEPLEKGYSH QLNYVMCFLM QGSRGTIPVL TWTHKSVDFF  
501 NMIDSKKITQ LPLVKAYKLQ SGASVVAGPR FTGGDIQCT ENGSAATIV  
551 TPDVSYSQY RARIHYASTS QITFTLSLDG APFNQYYFDK TINKGDTLTY  
601 NSFNLA SFST PFELSGNNLQ IGVTGLSAGD KYVIDKIEFI PVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

2 REFERENCES IN FILE CA (1907 TO DATE)

2 REFERENCES IN FILE CAPLUS (1907 TO DATE)

L15 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2004 ACS on STN

RN 112659-38-8 REGISTRY

CN Toxin, endo- (Bacillus thuringiensis tenebrionis parasporal crystal reduced) (9CI) (CA INDEX NAME)

FS PROTEIN SEQUENCE

SQL 594

SEQ 1 DNNTAELDSS TTKDVIQKGI SVVGDLGGV GFPPGGALVS FYTNFLNTIW  
51 PSEDPWKAFM EQVEALMDQK IADYAKNKAL AELQGLQNNV EDYVSALSSW  
101 QKNPVSSRNP HSQGRIRELF SQAESHFRNS MPSFAISGYE VLFLTTYAQA  
151 ANTHLFLKLD AQYGEWGY EKEDIAEFYK RQLKLTQEYT DHCVKWYNVG  
201 LDKLRGSSYE SWVNFNRYRR EMTLTVLDLI ALFPLYDVRL YPKEVKTELT  
251 RDVLTDPIVG VNNLRGYGT FSNENYIRKP HLFDYLHRI QFHTFRFQPGY  
301 YGNDSFNWS GNYVSTRPSI GSNDIITSPF YGNKSSEPQVQ NEFNGEKVY  
351 RAVANTNLAV WPSAVSGVT KVEFSQYNDQ TDEASTQTYD SKRNVGAVSW  
401 DSIDLPPET TDEPLEKGYSH QLNYVMCFL MQSGRGTPV LWTHKSVDFF  
451 NMIDSKKIT QPLVKAYKL QSGASVVAGP RFTGGDIQCT TENGSAATIV  
501 VTPDVSYSQ YRARIHYAST SQITFTLSLD GAPFNQYYFD KINKGDTLTY  
551 YNSFNLA SFS TPFELSGNNL QIGVTGLSAG DKYVIDKIEF IPVN

MF Unspecified

CI MAN

SR CA

LC STN Files: CA, CAPLUS, TOXCENTER

1 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES IN FILE CAPLUS (1907 TO DATE)



Set	Items	Description
S1	3029	THURINGIENSIS
S2	53940	TOXIN
S3	721	S1 AND S2
S4	56124	PROTEOL?
S5	72	S3 AND S4
S6	98699	FRAGMENT
S7	52	S3 AND S6
S8	43	S7 NOT S5

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- The conformational characteristics of the minimal toxic fragment of the delta-endotoxin from *Bacillus thuringiensis* berliner 1715 were examined by fluorescence and circular dichroism spectroscopy. This insecticidal protein, specifically toxic to lepidopteran species, was found to consist of two structural domains. Experimental evidence for this conclusion was provided by biphasic guanidine hydrochloride unfolding curves at different pH values and electrophoretic patterns of protease digest
- Two stable fragments of comparable molecular weight were obtained using four different broad specificity proteolytic enzymes. A secondary structure model was constructed using seven *B. thuringiensis* toxin sequences. These toxins were selected on the basis of their limited sequence homology and represent all known insecticidal specificities. Despite this divergence, a consensus secondary structure pattern was obtained, confirming the structural homology among the toxin
- The N-terminal halves of all toxins are predicted to be relatively rich in alpha-helix structure and the C-terminal parts to contain alternating beta-strand and coil structures. The latter seems characteristic for a beta-sheet conformation. Comparing model to the unfolding data obtained by circular dichroism, whose far UV signal gives a measure of the alpha-helix content allowed us to delineate the structural domains into the primary structure. Record Date Created: 19900221 Record Date Completed: 19900221
- 57/60 DIALOG(R)/File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.
- 08262992 PMID: 2549968 Proteolytic processing of a coleopteran-specific delta-endotoxin produced by *Bacillus thuringiensis* var. tenebrionis. Carroll J; Li J; Ellar D J Department of Biochemistry, University of Cambridge, U.K. Biochemical journal (ENGLAND) Jul 1 1989, 261 (1) p99-105, ISSN 0264-6021 Journal Code: 2984726R Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed
- Insecticidal protein delta-endotoxin crystals harvested from sporulated cultures of *Bacillus thuringiensis* var. tenebrionis contain a major polypeptide of 67 kDa and minor polypeptides of 73, 72, 55 and 46 kDa. During sporulation, only the 73 kDa polypeptide could be detected at stage I. The 67 kDa polypeptide was first detected at stage II and increased in concentration throughout the later stages of sporulation and after crystal release, with a concomitant decrease in the 73 kDa polypeptide. The change could be blocked by the addition of proteinase inhibitors. Trypsin or insect-gut-tract treatment of the delta-endotoxin crystals after solubilization resulted in a cleavage product of 55 kDa with asparagine-159 of the deduced amino acid sequence of the toxin [Hofte, Seurinck, van Houtven & Vaecq (1987) Nucleic Acids Res. 15, 71-83; Sekar, Thompson Maroney, Bookland & Adang (1987) Proc. Natl. Acad. Sci. U.S.A. 84, 7036-7040; McPherson, Penak, Fuchs, Marrone Lavrik & Fischhoff (1988) Biotechnology 6, 61-66] at the N-terminus. This polypeptide was found to be as toxic in vivo as native delta-endotoxin. Record Date Created: 19890927 Record Date Completed: 19890927
- 57/69 DIALOG(R)/File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.
- 07012872 PMID: 3909962 Protease activation of the entomotoxin protoxin of *Bacillus thuringiensis* subsp. kurstaki. Andrews R E; Bibilos M M; Bulla L A Applied and environmental microbiology (UNITED STATES) Oct 1985, 50 (4) p737-42, ISSN 0099-2240 Journal Code: 7605801 Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed



Two isolates of *Bacillus thuringiensis* subsp. *kurstaki* were examined which produced different levels of intracellular proteases. Although the crystals from both strains had comparable toxicity, one of the strains, LB1, had a strong polypeptide band at 68,000 molecular weight in the protein from the crystal; in the other, HD251, no such band was evident. When the intracellular proteases in both strains were measured, strain HD251 produced less than 10% of the proteolytic activity found in LB1. These proteases were primarily neutral metalloproteases, although low levels of other proteases were detected. In LB1, the synthesis of protease increased as the cells began to sporulate; however, in HD251, protease activity appeared much later in the sporulation cycle. The protease activity in strain LB1 was very high when the cells were making crystal toxin, whereas in HD251 reduced proteolytic activity was present during crystal toxin synthesis. The insecticidal toxin (molecular weight, 68,000) from both strains could be prepared by cleaving the protoxin (molecular weight, 135,000) with trypsin, followed by ion-exchange chromatography. The procedure described gave quantitative recovery of toxic activity, and approximately half of the total protein was recovered. Calculations show that these results correspond to stoichiometric conversion of protoxin to insecticidal toxin. The toxicities of whole crystals, soluble crystal protein, and purified toxin from both strains were comparable. Record Date Created: 19860130 Record Date Completed: 19860130

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07102416 PMID: 3009187

Specificity of *Bacillus thuringiensis* var. *colimae* insecticidal delta-endotoxin is determined by differential proteolytic processing of the protoxin by larval gut proteases.

Haider M Z; Knowles B H; Ellar D J  
European journal of biochemistry / FEBS (GERMANY, WEST) May 2 1986; 156 (3): p531-40, ISSN 0014-2956 Journal Code: 0107600 Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed  
The native crystal delta-endotoxin produced by *Bacillus thuringiensis* var. *colimae*, serotype 21, is toxic to both lepidopteran (Pieris brassicae) and dipteran (*Aedes aegypti*) larvae. Solubilization of the crystal delta-endotoxin in alkaline reducing conditions and activation with trypsin and gut extracts from susceptible insects yielded a preparation whose toxicity could be assayed *in vitro* against a range of insect cell lines. After activation with *Aedes aegypti* gut extract the preparation was toxic to all of the mosquito cell lines but only one lepidopteran line (*Spodoptera frugiperda*), whereas an activated preparation produced by treatment with *P. brassicae* gut enzymes or trypsin was toxic only to lepidopteran cell lines. These *in vitro* results were paralleled by the results of *in vivo* bioassays. Gel electrophoretic analysis of the products of these different activation regimes suggested that a 130-kDa protoxin in the native crystal is converted to a 55-kDa lepidopteran-specific toxin by trypsin or *P. brassicae* enzymes and to a 52-kDa dipteran toxin by *A. aegypti* enzymes. Two-step activation of the 130-kDa protoxin by successive treatment with trypsin and *A. aegypti* enzymes further suggested that the 52-kDa dipteran toxin is derived from the 55-kDa lepidopteran toxin by enzymes specific to the mosquito gut. Confirmation of this suggestion was obtained by peptide mapping of these two polypeptides. The native crystal 130 kDa delta-endotoxin and the two insect-specific toxins all cross-reacted with antiserum to *B. thuringiensis* var. *kurstaki* P1 lepidopteran toxin. Preincubation of the two activated colimae toxins with P1 antiserum neutralized their cytotoxicity to both lepidopteran and dipteran cell lines. Record Date Created: 19860618 Record Date Completed: 19860618

57/770 DIALOG(R)/File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.

06813876 PMID: 3888979

Delineation of a toxin-encoding segment of a *Bacillus thuringiensis* crystal protein gene.

Schnepf H E; Whiteley H R  
Journal of biological chemistry (UNITED STATES) May 25 1985; 260 (10): p6273-80, ISSN 0021-9258 Journal Code: 2985121R Contract/Grant No.: GM-20784; GM; NIGMS; GM-26100; GM; NIGMS; K5-GM-442; GM; NIGMS Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed

Crystals of *Bacillus thuringiensis* subsp. *kurstaki* HD-1-Dipel contain a Mr 134,000 protoxin which can be cleaved by proteolysis to a peptide of Mr approximately 70,000; this peptide is lethal to lepidopteran larvae. We have analyzed the peptides produced by recombinant *Escherichia coli* strains bearing deletions and fusions of the protoxin gene in order to delineate the portion of the gene which encodes the toxic peptide. The recombinant strains produced the toxic peptide as well as larger peptides whose size was related to the length of the deleted gene. The results indicate that the amino-terminal 55% of the protoxin protein is sufficient for toxicity. While two different gene fusions to the 10th codon allowed the synthesis of toxic polypeptides, fusions to the 50th codon did not. 3' end deletions up to the 645th codon allowed synthesis of the toxic peptide whereas a deletion to the 603rd codon yielded a non-toxic peptide. Some of the 5' and 3' end alterations to the gene caused changes in the proteolytic cleavage patterns of the polypeptides synthesized by *E. coli*, suggesting that the alterations led to conformational changes in the proteins. The presence of different 3' end segments affected the levels of synthesis of the altered crystal proteins. Record Date Created: 19850627 Record Date Completed: 19850627

57/771 DIALOG(R)/File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.  
06465876 PMID: 6321329

Specificity of cultured insect tissue cells for bioassay of entomocidal protein from *Bacillus thuringiensis*.  
Johnson D E; Davidson L I

*In vitro* (UNITED STATES) Jan 1984; 20 (1): p66-70, ISSN 0073-5655 Journal Code: 00637333 Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed  
Cultured tissue cells from lepidopteran and dipteran sources displayed an order-specific response to entomocidal protein from crystals of *Bacillus thuringiensis*. Protein isolated from crystals of *B. thuringiensis* subsp. *kurstaki* (Manduca sexta) but was against cells of the spruce budworm (*Choristoneura fumiferana*) and the tobacco hornworm (*Manduca sexta*) but was inactive against both mosquito cell lines tested (*Aedes aegypti* and *Anopheles gambiae*). Conversely, protein from inclusions of *B. thuringiensis* subsp. *israelensis* was fully active only against the mosquito cell lines but displayed reduced (four- to seven-fold) toxicity for the lepidopteran cell lines. One exception to this pattern of specificity was observed with *Plodia interpunctella* cell line, which failed to respond to either crystal protein preparation. The moth toxin was stable at degrees C for months, whereas the mosquito toxin was susceptible to proteolytic degradation and was unstable for periods longer than 2 wk. Record Date Created: 19840419 Record Date Completed: 19840419

57/772 DIALOG(R)/File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.  
05398675 PMID: 7420053

Purification of the insecticidal toxin in crystals of *Bacillus thuringiensis*.

Lilley M; Ruffell R N; Somerville H J

Journal of general microbiology (ENGLAND) May 1980; 118 (Pt 1): p1-11, ISSN 0022-1287 Journal Code: 0375371 Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed

Crystals were purified from four serotypes of the insect pathogen *Bacillus thuringiensis*. Crystals from these serotypes were similar in amino acid and N-terminal analyses, but differed in their toxicity to two species of Lepidoptera and in their immunological properties. Toxic polypeptides were obtained following trypsin digestion of solutions of the crystals. In two strains (serotypes 3 and 9) this fraction contained only one polypeptide. Similar results were obtained when dissolved crystals were digested with other proteolytic enzymes or with gut contents from Pieris brassicae. The trypsin-resistant polypeptide was further purified by gel and ion-exchange chromatography and had a molecular weight of about 70,000, estimated by gel chromatography and gel electrophoresis. No evidence was obtained for a toxin of lower molecular weight. This purified toxin accounted for most, if not all, of the toxic activity originally present in the crystal solution and was active by injection and ingestion. The purified toxic fraction from serotype 1 appeared to contain two polypeptides, one of which corresponded to that found with serotypes 3 and 9. There were no major differences in the composition of crystals from different serotypes of *B. thuringiensis* and it is concluded that the trypsin-resistant polypeptide represents the active insecticidal toxin of the crystal. Record Date Created: 19801218 Record Date Completed: 19801218

8/6/1 15683588 PMID: 14668140

Ion channels formed in planar lipid bilayers by the dipteran-specific Cry4B *Bacillus thuringiensis* toxin and its alpha1-alpha8 fragment. Ja Feb 2004

8/6/2 15013850 PMID: 12555398

Cloning and expression of the binary toxin genes of *Bacillus sphaericus* C3-41 in a crystal minus *B. thuringiensis* subsp. *israelensis* Feb 199

8/6/3 14510132 PMID: 10508095

Production of chymotrypsin-resistant *Bacillus thuringiensis* Cry2Aa1 delta-endotoxin by protein engineering. Oct 1999

8/6/4 14485285 PMID: 10481060

Amino acid substitution in alpha-helix 7 of Cry1Ac delta-endotoxin of *Bacillus thuringiensis* leads to enhanced toxicity to *Helicoverpa armigera* Hubner. Sep 17 1999

8/6/5 14373264 PMID: 10366728

*Bacillus thuringiensis* insecticidal Cry1Aa toxin binds to a highly conserved region of aminopeptidase N in the host insect leading to its evolutionary success. Jun 15 1999

8/6/6 14060546 PMID: 9758818

The introduction into *Bacillus sphaericus* of the *Bacillus thuringiensis* subsp. *medellin* Cry1Ab1 gene results in higher susceptibility of resistant mosquito larva populations to *B. sphaericus*. Oct 1998

8/6/7 14040117 PMID: 9739466

Biochemical characterization of the third domain from *Bacillus thuringiensis* Cry1A toxins. Aug 1998

8/6/8 13648195 PMID: 9342226

Cloning and characterization of *Manduca sexta* and *Plutella xylostella* midgut aminopeptidase N enzymes related to *Bacillus thuringiensis* toxin-binding proteins. Sep 15 1997

8/6/9 13639101 PMID: 9332588

Cloning, expression and toxicity of a mosquitoicidal toxin gene of *Bacillus thuringiensis* subsp. *medellin*. Mar-Apr 1997

8/6/10 13626264 PMID: 9315709

Isolated domain II and III from the *Bacillus thuringiensis* Cry1Ab delta-endotoxin binds to lepidopteran midgut membranes. Sep 8 1997



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Thermodynamic analysis of domain organization of *Bacillus thuringiensis* toxins [ Termodinamicheeskii analiz domennoi organizatsii toksinov *Bacillus thuringiensis*. Dec 1996
- 8/6/12 12893251 PMID: 8572715  
A *Bacillus thuringiensis* insecticidal crystal protein with a high activity against members of the family Noctuidae. Jan 1996
- 8/6/13 12605248 PMID: 7722342  
The insecticidal CryB crystal protein of *Bacillus thuringiensis* ssp. *thuringiensis* has dual specificity to coleopteran and lepidopteran larvae. Mar 1995
- 8/6/14 12357447 PMID: 12729731  
Expression in *Pichia pastoris* and purification of a membrane-acting immunotoxin based on a synthetic gene coding for the *Bacillus thuringiensis* Cyt2Aa1 toxin. May 2003
- 8/6/15 12168626 PMID: 12502392  
Enterotoxigenicity and cytotoxicity of *Bacillus thuringiensis* strains and development of a process for Cry1Ac production. Jan 1 2003
- 8/6/16 12123166 PMID: 12452809  
Heterologous expression of cry2 gene from a local strain of *Bacillus thuringiensis* isolated in Nigeria. Dec 2002
- 8/6/17 12036661 PMID: 12357073  
Cloning and characterization of an insecticidal crystal protein gene from *Bacillus thuringiensis* subspecies *kenyae*. Apr 2002
- 8/6/18 11998402 PMID: 12213239  
Cry1A toxins of *Bacillus thuringiensis* bind specifically to a region adjacent to the membrane-proximal extracellular domain of BT-R(1) in *Manduca sexta*: involvement of a cadherin in the entomopathogenicity of *Bacillus thuringiensis*. Sep 2002
- 8/6/19 11639598 PMID: 11815850  
Physical mapping of the *Bacillus thuringiensis* subsp. *kurstaki* and *alesti* chromosomes. Feb 2002
- 8/6/20 11474687 PMID: 11583928  
Processing of Cry1Ab delta-endotoxin from *Bacillus thuringiensis* by *Manduca sexta* and *Spodoptera frugiperda* midgut proteases: role in protoxin activation and toxin inactivation. Nov 1 2001
- 8/6/21 10891750 PMID: 11023737  
Characterization of a *Bacillus thuringiensis* delta-endotoxin which is toxic to insects in three orders. Aug 2000
- 8/6/22 10855333 PMID: 10985018  
Comparative insecticidal properties of two nucleopolyhedrovirus vectors encoding a similar toxin gene chimera. Aug 2000
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*Bacillus thuringiensis* Cry1Aa toxin-binding region of *Bombyx mori* aminopeptidase N. Dec 17 1999
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Genome stability of *Bacillus thuringiensis* subsp. *israelensis* isolates. Jan 2000
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Insect-resistant chrysanthemum calluses by introduction of a *Bacillus thuringiensis* crystal protein gene. May 1993
- 8/6/26 09704360 PMID: 8491716  
Full expression of the cryIIIA toxin gene of *Bacillus thuringiensis* requires a distant upstream DNA sequence affecting transcription. May 1993
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Cloning of a novel cryIc-type gene from a strain of *Bacillus thuringiensis* subsp. *galleriae*. Apr 1993
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Genomic amplification and expression of delta-endotoxin fragment of *Bacillus thuringiensis*. Sep 16 1992
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Expansion of insecticidal host range of *Bacillus thuringiensis* by in vivo genetic recombination. Apr 1992
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Insecticidal properties of a crystal protein gene product isolated from *Bacillus thuringiensis* subsp. *kenyae*. Feb 1992
- 8/6/31 08813686 PMID: 1847865  
Two structural domains as a general fold of the toxic fragment of the *Bacillus thuringiensis* delta-endotoxins. Feb 14 1991
- 8/6/32 08618820 PMID: 2200339  
Molecular cloning of the 130-kilodalton mosquitoicidal delta-endotoxin gene of *Bacillus thuringiensis* subsp. *israelensis* in *Bacillus sphaericus*. Jun 1990
- 8/6/33 08491852 PMID: 2323547  
Heterologous expression of a mutated toxin gene from *Bacillus thuringiensis* subsp. *tenebrionis*. Jan 1 1990
- 8/6/34 08267548 PMID: 2550328  
Gene dosage effect on the expression of the delta-endotoxin genes of *Bacillus thuringiensis* subsp. *kurstaki* in *Bacillus subtilis* and *Bacillus megaterium*. Jun 30 1989
- 8/6/35 08014170 PMID: 2536653  
Involvement of Tr-4430 in transfer of *Bacillus anthracis* plasmids mediated by *Bacillus thuringiensis* plasmid pXO12. Jan 1989
- 8/6/36 07793333 PMID: 2897850  
The mosquito larvicidal activity of 130 kDa delta-endotoxin of *Bacillus thuringiensis* var. *israelensis* resides in the 72 kDa amino-terminal fragment. May 31 1988
- 8/6/37 07739001 PMID: 2833395  
Common features of *Bacillus thuringiensis* toxins specific for Diptera and Lepidoptera. Apr 5 1988
- 8/6/38 07506411 PMID: 3038691  
Cloning and heterologous expression of an insecticidal delta-endotoxin gene from *Bacillus thuringiensis* var. *aizawai* IC1 toxic to both lepidoptera and diptera. 1987
- 8/6/39 07402365 PMID: 3031425  
Characterization of the genes encoding the haemolytic toxin and the mosquitoicidal delta-endotoxin of *Bacillus thuringiensis* *israelensis*. D 1986
- 8/6/40 07331004 PMID: 3026371  
Purification and characterization of the active fragment from *Bacillus thuringiensis* delta-toxin. Nov 26 1986
- 8/6/41 07317812 PMID: 3025452  
*Bacillus thuringiensis* var. *israelensis* delta-endotoxin. Nucleotide sequence and characterization of the transcripts in *Bacillus thuringiensis* and *Escherichia coli*. Sep 5 1986
- 8/6/42 07158027 PMID: 3013729  
Cloning and expression of the lepidopteran toxin produced by *Bacillus thuringiensis* var. *thuringiensis* in *Escherichia coli*. 1986
- 8/6/43 06615870 PMID: 6090216  
Cloning and expression in *Escherichia coli* of the insecticidal delta-endotoxin gene of *Bacillus thuringiensis* var. *israelensis*. Oct 1 1984
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07317812 PMID: 3025452  
*Bacillus thuringiensis* var. *israelensis* delta-endotoxin. Nucleotide sequence and characterization of the transcripts in *Bacillus thuringiensis* and *Escherichia coli*.  
Ward E S; Ellar D J  
Journal of molecular biology (ENGLAND) Sep 5 1986, 191 (1) p1-11, ISSN 0022-2836 Journal Code: 2985088R  
Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed  
The nucleotide sequence of a 1408 base-pair DNA fragment encoding the insecticidal 27,340 Mr delta-endotoxin of *Bacillus thuringiensis* var. *israelensis* has been determined by analysis of a recombinant plasmid from *Escherichia coli*. The hydrophathy plot of the protein shows it to be highly hydrophobic, consistent with a postulated cytolytic mechanism of action for the toxin. In addition, the delta-endotoxin transcriptional start points that are used in *B. thuringiensis* and an *E. coli* recombinant have been determined. In *B. thuringiensis* var. *israelensis*, transcription initiates from a single start point, and gene-specific transcripts are not observed before stage II of sporulation. This is the stage at which delta-endotoxin antigen is first detected, indicating that control of expression is primarily at the transcriptional level for this protein. Analysis of gene-specific transcription in *E. coli* indicates that at least three start points are utilized in this organism. Interestingly, the highest level of delta-endotoxin mRNA is seen during mid-exponential growth of *E. coli* and the level appears to decrease as the cells enter the stationary phase of growth. Record Date Created: 19870123 Record Date Completed: 19870123
- 8/7/42 DIALOG(R)/File 155: MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.  
07158027 PMID: 3013729  
Cloning and expression of the lepidopteran toxin produced by *Bacillus thuringiensis* var. *thuringiensis* in *Escherichia coli*.  
Honigman A; Nedjar-Pazemli G; Yawetz A; Oron U; Schuster S; Broza M; Sneh B  
Gene (NETHERLANDS) 1986, 42 (1) p69-77, ISSN 0378-1119 Journal Code: 7706761 Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed  
The *Bacillus thuringiensis* var. *thuringiensis* strain 3A produces a proteinaceous parasporal crystal toxic to larvae of a variety of lepidopteran pests including *Spodoptera littoralis* (Egyptian cotton leaf worm), *Heliothis zea*, *H. virescens* and *Boarmia selenaria*. By cloning of individual plasmids of *B. thuringiensis* in *Escherichia coli*, we localized a gene coding for the delta-endotoxin on the *B. thuringiensis* plasmid of about 17 kb designated pTN4. Following partial digestion of the



thuringiensis plasmid pTM4 and cloning into the E. coli pACYC184 plasmid three clones were isolated in which toxin production was detected. One of these hybrid plasmids pTNG43 carried a 1.7-kb insert that hybridized to the 14-kb BamHI DNA fragments of B. thuringiensis var. thuringiensis strains 3A and berliner 1715. This BamHI DNA fragment of strain berliner 1715 has been shown to contain the gene that codes for the toxic protein of the crystal (Klier et al., 1982). No homologous sequences have been found between pTNG33 and the DNA of B. thuringiensis var. entomocidus strain 24, which exhibited insecticidal activity against S. littoralis similar to that of strain 3A. Record Date Created: 19880725 Record Date Completed: 19860725

87/43 DIALOG(R)File 155:MEDLINE(R) (c) format only 2004 The Dialog Corp. All rts. reserv.

06615870 PMID: 6090216

Cloning and expression in Escherichia coli of the insecticidal delta-endotoxin gene of Bacillus thuringiensis var.

israelensis. Ward E S; Elar D J; Todd J A

FEBS letters (NETHERLANDS) Oct 1 1984, 175 (2), p377-82, ISSN 0014-5793 Journal Code: 0155157

Document type: Journal Article Languages: ENGLISH Main Citation Owner: NLM Record type: Completed

Recombinant plasmids containing the mosquitoicidal delta-endotoxin gene were constructed by inserting HindIII

fragments of the Bacillus thuringiensis var. israelensis 72.75 Md plasmid in to the Escherichia coli vector pUC12. Two

recombinants producing the 26 000 Da delta-endotoxin (pIP173 and pIP174) were identified by screening clones in an E. coli in

vitro transcription-translation system. Both recombinants were 12.4 kb chimaeric plasmids comprising pUC12 and a common

9.7 kb HindIII fragment of the B. thuringiensis plasmid. The 26 000 Da polypeptide synthesis in vivo from pIP174

transformed into E. coli JM101 was lethal to mosquito larvae and cytotoxic to mosquito cells in vitro. The biological

authenticity of the cloned product was further confirmed by demonstrating that the cytotoxicity of the polypeptide was

neutralised by antiserum to the authentic delta-endotoxin or by preincubation with excess toxin receptor. Transcription of

the recombinant delta-endotoxin gene in E. coli appears to utilise a Bacillus promoter sequence(s) rather than the pUC12 beta-

galactosidase promoter. Record Date Created: 19841119 Record Date Completed: 19841119

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Set Items Description

S1 8862 THURINGIENSIS

S2 141509 TOXIN

S3 1859 S1 AND S2

S4 67863 PROTEOL?

S5 94 S3 AND S4

S6 113606 FRAGMENT

S7 84 S3 AND S6

S8 73 S7 NOT S5

5/6/1

0014626028 BIOSIS NO.: 2003000578705

Role of proteolysis in determining potency of Bacillus thuringiensis

Cry1Ac delta-endotoxin.

2000

5/6/2 0014531207 BIOSIS NO.: 200300488864

The mode of action of the Bacillus thuringiensis vegetative insecticidal protein Vip3A differs from that of Cry1Ab delta-endotoxin. 2003

5/6/3 0014524885 BIOSIS NO.: 200300478840

Mutation of the hydrophobic residue on helix alpha5 of the Bacillus thuringiensis Cry4B affects structural stability. 2003

5/6/4 0014447768 BIOSIS NO.: 200300405487

Activity of free and clay-bound insecticidal proteins from Bacillus thuringiensis subsp. israelensis against the mosquito Culex pipiens. 2003

5/6/5 0014194705 BIOSIS NO.: 200300153424

Characterization and comparison of midgut proteases of Bacillus thuringiensis susceptible and resistant diamondback moth (Plutellidae: Lepidoptera). 2003

5/6/6 0014142486 BIOSIS NO.: 200300101205

Mode of action of Cry toxins from Bacillus thuringiensis. ] ORIGINAL LANGUAGE TITLE: Mecanismo de accion de las toxinas Cry de Bacillus

thuringiensis. 2002

5/6/7 0013695372 BIOSIS NO.: 200200488883

Production of delta-endotoxin by Bacillus thuringiensis subsp kurstaki and overcoming of catabolite repression by using highly concentrated

gruel and fish meal media in 2- and 20-dm3 fermenters 2002

5/6/8 0013834530 BIOSIS NO.: 200200428041

N-terminal activation is an essential early step in the mechanism of action of the Bacillus thuringiensis Cry1Ac insecticidal toxin 2002

5/6/9 0013811889 BIOSIS NO.: 200200405400

Characterization of the proteolytic enzymes in the midgut of the European Cockchafer, Melolontha melolontha (Coleoptera: Scarabaeidae) 20

5/6/10 0013798038 BIOSIS NO.: 200200391549

Interaction of 65- and 62-kD proteins from the apical membranes of the Aedes aegypti larvae midgut epithelium with Cry4B and Cry11A ndolo

of Bacillus thuringiensis 2002

5/6/11 0013721379 BIOSIS NO.: 200200314890

Changes in protease activity and Cry3Aa toxin binding in the Colorado potato beetle: Implications for insect resistance to Bacillus thuringiensis

toxins 2002

5/6/12 0013557352 BIOSIS NO.: 200200250863

Cadherin-like receptor binding facilitates proteolytic cleavage of helix alpha-1 in domain I and oligomer pre-pore formation of Bacillus

thuringiensis Cry1Ab toxin 2002

5/6/13 0013529422 BIOSIS NO.: 200200222933

Colorado potato beetle resistance to the Cry3A toxin of Bacillus thuringiensis subsp. Tenebrionis 2001

5/6/14 0013540198 BIOSIS NO.: 200200133709

Bacillus thuringiensis - An insecticide. ] ORIGINAL LANGUAGE TITLE: Bacillus thuringiensis : W zwalczaniu owadow 2001

5/6/15 0013395843 BIOSIS NO.: 200100567682

Transcriptional activator PtcR regulate the expression of multiple genes in Bacillus cereus 2001

5/6/16 0013314287 BIOSIS NO.: 200100466126

Role of interdomain salt bridges in the pore-forming ability of the Bacillus thuringiensis toxins Cry1Aa and Cry1Ac 2001

5/6/17 0013074836 BIOSIS NO.: 200100246675

Specific cleavage of the Cry1Ab toxin receptor BT-R1 in the midgut epithelium of Manduca Sextia 2001

5/6/18 0013072144 BIOSIS NO.: 200100243983

Redesign of an interhelical loop of the Bacillus thuringiensis Cry4B delta-endotoxin for proteolytic cleavage 2001

5/6/19 0012783207 BIOSIS NO.: 200000501520

Membrane pore architecture of a cytolytic toxin from Bacillus thuringiensis 2000

5/6/20 0012745539 BIOSIS NO.: 200000463852

Proteolytic processing of the Cyt1Ab1 toxin produced by Bacillus thuringiensis subsp. Medellin 2000

5/6/21 0012577881 BIOSIS NO.: 200000395194

Activation pattern and toxicity of the Cry11Bb1 toxin of Bacillus thuringiensis subsp. Medellin 2000

5/6/22 0012536933 BIOSIS NO.: 200000355246

The interactions between soybean trypsin inhibitor and delta-endotoxin of Bacillus thuringiensis in Helicoverpa armigera larva 2000

5/6/23 0012585368 BIOSIS NO.: 200000304681

Methanol-induced conformational changes in delta-endotoxin from Bacillus thuringiensis var. tenebrionis 2000

5/6/24 0012403218 BIOSIS NO.: 200000121531

Identification and purification of the 69-kDa intracellular protease involved in the proteolytic processing of the crystal delta-endotoxin of Bacil

thuringiensis subsp. Tenebrionis 2000

5/6/25 0012293594 BIOSIS NO.: 200000011907

Activation and fragmentation of Bacillus thuringiensis delta-endotoxin by high concentrations of proteolytic enzymes 1999

5/6/26 0012193125 BIOSIS NO.: 199900452785

Resistance to Bacillus thuringiensis Cry1Ac toxin in three strains of Heliothis virescens: Proteolytic and SEM study of the larval midgut 1999

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Intragastric and intraperitoneal administration of Cry1Ac protoxin from Bacillus thuringiensis induces systemic and mucosal antibody response

in mice 1999

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Biochemical characterization of Bacillus thuringiensis cytolytic toxins in association with a phospholipid bilayer 1999

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Proteolysis of *Bacillus thuringiensis* subspecies *kurstaki* endotoxin with midgut proteases of some important lepidopterous species 1998

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Involvement of an endogenous metalloprotease in the activation of protoxin in *Bacillus thuringiensis* subsp. *kurstaki* 1997

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Correction of PREVIEWS 99342806. Mapping of the entomocidal fragment of *Spodoptera*-specific *Bacillus thuringiensis* toxin CryIC. Correction of author name from Z. Konez-Kalman. Erratum published in Molecular and General Genetics Vol. 253, Iss. 6, 1997, p. 777-1996

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The effect of *Bacillus thuringiensis* M-toxin on trematode cercariae 1996

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Digestion of delta-endotoxin by gut proteases may explain reduced sensitivity of advanced instar larvae of *Spodoptera littoralis* to CryIC 1996

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Proteolytic processing of *Bacillus thuringiensis* CryIIIA toxin and specific binding to brush-border membrane vesicles of *Lepidodarsa decemlineata* (Colorado potato beetle) 1996

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Efficient synthesis of mosquitoicidal toxins in *Aspicacaulis excentricus* demonstrates potential of gram-negative bacteria in mosquito control 1996

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Structure of the mosquitoicidal delta-endotoxin CytB from *Bacillus thuringiensis* sp. *kyushuensis* and implications for membrane pore formation 1996

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Differences in the midgut proteolytic activity of two *Heliothis virescens* strains, one susceptible and one resistant to *Bacillus thuringiensis* toxins 1996

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Mosquitoicidal activity of the CryIC delta-endotoxin from *Bacillus thuringiensis* subsp. *Aizawai* 1996

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Functional significance of loops in the receptor binding domain of *Bacillus thuringiensis* CryIIIA delta-endotoxin 1996

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The use of *Bacillus thuringiensis* in crop protection and the development of pest resistance 1995

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Maximal toxicity of cloned CytA delta-endotoxin from *Bacillus thuringiensis* subsp. *israelensis* requires proteolytic processing from both the N and C-termini 1995

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Mapping and characterization of the entomocidal domain of the *Bacillus thuringiensis* CryIA(b) protoxin 1995

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Production of multiple delta-endotoxins by *Bacillus thuringiensis*: delta-endotoxins produced by strains of the subspecies *galleriae* and *wuhanensis* 1994

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Expression of the insecticidal crystal protein gene from a Gram-positive *Bacillus thuringiensis* in a Gram-negative *Pseudomonas fluorescens* mediated by protoplast fusion 1994

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Intracellular proteolysis and limited diversity of the *Bacillus thuringiensis* CryIA family of the insecticidal crystal proteins 1994

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Characterization of larvicidal toxin protein from *Bacillus thuringiensis* serovar *japonensis* strain *Buibui* specific for scarabaeid beetles 1994

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Toxicity of activated CryI proteins from *Bacillus thuringiensis* to six forest Lepidoptera and *Bombyx mori* 1993

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Structural stability of *Bacillus thuringiensis* delta-endotoxin homolog-scanning mutants determined by susceptibility to proteases 1993

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An analysis of *Bacillus thuringiensis* delta-endotoxin action on insect-midgut-membrane permeability using a light-scattering assay 1993

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A specific binding protein from *Manduca sexta* for the insecticidal toxin of *Bacillus thuringiensis* ssp. *Berliner* 1993

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In vitro and in vivo proteolysis of the *Bacillus thuringiensis* ssp. *israelensis* CryIVD protein by *Culex quinquefasciatus* larval midgut protease 1993

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PROTEOLYTIC PROCESSING OF DELTA ENDOTOXIN OF *BACILLUS THURINGIENSIS* -VAR-KURSTAKI HD-1 IN INSENSITIVE INSECT SPODOPTERA-LITURA UNUSUAL PROTEOLYSIS IN THE PRESENCE OF SODIUM DODECYL SULFATE 1992

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MODE OF ACTION OF *BACILLUS THURINGIENSIS* TOXIN CRYIIIA AN ANALYSIS OF TOXICITY IN LEPTINOTARSA-DECEMLINEATA SAY AND DIABROTICA-UNDECIMPUNCTATA-HOWARDI BARBER 1992

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CYTOTOXICITY OF A CLONED *BACILLUS THURINGIENSIS* -SSP-ISRAELENSIS CRYIVB TOXIN TO AN AEDES-AEGYPTI CELL LINE 19

5/6/71 0007885743 BIOSIS NO.: 199192131514



- THE TOXIC MOIETY OF THE BACILLUS- THURINGIENSIS PROTOXIN UNDERGOES A CONFORMATIONAL CHANGE UPON ACTIVATION 1991
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FOLDING AND UNFOLDING OF THE PROTOXIN FROM BACILLUS- THURINGIENSIS EVIDENCE THAT THE TOXIC MOIETY IS PRESENT IN AN ACTIVE CONFORMATION 1990
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PROTEOLYTIC PROCESSING OF A COLEOPTERAN-SPECIFIC DELTA ENDOTOXIN PRODUCED BY BACILLUS- THURINGIENSIS -VAR- TENEBRIONIS 1989
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PURIFICATION AND PROPERTIES OF A 28-KILODALTON HEMOLYTIC AND MOSQUITOCIDAL PROTEIN TOXIN OF BACILLUS- THURINGIENSIS -SSP-DARMSTADTENSIS 73-E10-2 1989
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DIFFERENTIAL SPECIFICITY OF TWO INSECTICIDAL TOXINS FROM BACILLUS- THURINGIENSIS -VAR-AIZAWAI 1988
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SPECIFICITY OF BACILLUS- THURINGIENSIS DELTA ENDOTOXINS IS CORRELATED WITH THE PRESENCE OF HIGH-AFFINITY BINDING SITES IN THE BRUSH BORDER MEMBRANE OF TARGET INSECT MIDGUTS 1988
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HOST SPECIFICITY OF THE BACILLUS- THURINGIENSIS DELTA ENDOTOXIN TOWARD LEPIDOPTERAN SPECIES SPODOPTERA-LITTORALIS BDV. AND PIERIS-BRASSICAE L 1987
- 5/6/86 0005215711 BIOSIS NO.: 198682062098  
SPECIFICITY OF BACILLUS- THURINGIENSIS -VAR-COLMERI INSECTICIDAL DELTA ENDOTOXIN IS DETERMINED BY DIFFERENTIAL PROTEOLYTIC PROCESSING OF THE PROTOXIN BY LARVAL GUT PROTEASES 1986
- 5/6/87 0005083756 BIOSIS NO.: 198681002647  
PROTEASE ACTIVATION OF THE ENTOMOCIDAL PROTOXIN OF BACILLUS- THURINGIENSIS -SSP-KURSTAKI 1985
- 5/6/88 0004740233 BIOSIS NO.: 198580049128  
DELINEATION OF A TOXIN -ENCODING SEGMENT OF A BACILLUS- THURINGIENSIS CRYSTAL PROTEIN GENE 1985
- 5/6/89 0004513711 BIOSIS NO.: 198529042610  
BIOSYNTHESIS OF THE INSECTICIDAL TOXIN FROM BACILLUS- THURINGIENSIS -SSP-ISRAELENSIS 1985
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DELINEATION OF A TOXIN -ENCODING SEGMENT OF A BACILLUS- THURINGIENSIS CRYSTAL PROTEIN GENE
- DISSOLUTION AND DEGRADATION OF BACILLUS- THURINGIENSIS DELTA ENDO TOXIN BY GUT JUICE PROTEASE OF THE SILKWORM BOMBYX-MORI 1983
- 5/6/91 0003883478 BIOSIS NO.: 198375067421  
THE MAIN FEATURES OF BACILLUS- THURINGIENSIS DELTA ENDO TOXIN MOLECULAR STRUCTURE 1982
- 5/6/92 0003025269 BIOSIS NO.: 198070056756  
PURIFICATION OF THE INSECTICIDAL TOXIN IN CRYSTALS OF BACILLUS- THURINGIENSIS 1980
- 5/6/93 0002286900 BIOSIS NO.: 197815004387  
SOLUBLE GLYCO PROTEIN INSECT TOXIN FROM THE SPORE COAT OF BACILLUS- THURINGIENSIS 1978
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FURTHER OBSERVATIONS ON THE MODE OF ACTION OF BACILLUS- THURINGIENSIS ON PAPILO-DEMOLEUS AND SPODOPTERA-LITURA 1976
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0005215711 BIOSIS NO.: 198682062098  
SPECIFICITY OF BACILLUS- THURINGIENSIS -VAR-COLMERI INSECTICIDAL DELTA ENDOTOXIN IS DETERMINED BY DIFFERENTIAL PROTEOLYTIC PROCESSING OF THE PROTOXIN BY LARVAL GUT PROTEASES  
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JOURNAL: European Journal of Biochemistry 156 (3): p531-540 1986 ISSN: 0014-2956 DOCUMENT TYPE: Article  
RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: The native crystal .delta.-endotoxin produced by *Bacillus thuringiensis* var. *colmeri*, serotype 21, is toxic to both lepidopteran (Pieris brassicae) and dipteran (*Aedes aegypti*) larvae. Solubilization of the crystal .delta.-endotoxin in alkaline reducing conditions and activation with trypsin and gut extracts from susceptible insects yielded a preparation whose toxicity could be assayed in vitro against a range of insect cell lines. After activation with *Aedes aegypti* gut extract the preparation was toxic to all of the mosquito cell lines but only one lepidopteran line (*Spodoptera frugiperda*), whereas an activated preparation produced by treatment with *P. brassicae* gut enzymes or trypsin was toxic only to lepidopteran cell lines. These in vitro results were paralleled by the results of in vivo bioassays. Gel electrophoretic analysis of the products of these different activation regimes suggested that a 130-kDa protoxin in the native crystal is converted to a 55-kDa lepidopteran-specific toxin by trypsin or *P. brassicae* enzymes and to a 52-kDa dipteran toxin by *A. aegypti* enzymes. Two-step activation of the 130-kDa protoxin by successive treatment with trypsin and *A. aegypti* enzymes further suggested that the 52-kDa dipteran toxin is derived from the 55-kDa lepidopteran toxin by enzymes specific to the mosquito gut. Confirmation of this suggestion was obtained by peptide mapping of these two polypeptides. The native crystal 130 kDa .delta.-endotoxin and the two insect-specific toxins all cross-reacted with antiserum to *B. thuringiensis* var. *kurstaki* P1 lepidopteran toxin. Preincubation of the two activated colmeri toxin with P1 antiserum neutralized their cytotoxicity to both lepidopteran and dipteran cell lines.
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0005038756 BIOSIS NO.: 198681002647  
PROTEASE ACTIVATION OF THE ENTOMOCIDAL PROTOXIN OF BACILLUS- THURINGIENSIS -SSP-KURSTAKI  
AUTHOR: ANDREWS R E JR (Reprint); BIBLOS M M; BULLA L A JR  
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JOURNAL: Applied and Environmental Microbiology 50 (4): p737-742 1985 ISSN: 0099-2240 DOCUMENT TYPE: Article  
RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: Two isolates of *Bacillus thuringiensis* subsp. *kurstaki* were examined which produced different levels of intracellular proteases. Although the crystals from both strains had comparable toxicity, one of the strains, LB1, had a strong polypeptide band at 68,000 molecular weight in the protein from the crystals; in the other, HD251, no such band was evident. When the intracellular proteases in both strains were measured, strain HD251 produced less than 10% of the proteolytic activity found in LB1. These proteases were primarily neutral metalloproteases, although low levels of other proteases were detected. In LB1, the synthesis of protease increased as the cells began to sporulate; however, in HD251, protease activity appeared much later in the sporulation cycle. The protease activity in strain LB1 was very high when the cells were making crystal toxin, whereas in HD251 reduced proteolytic activity was present during crystal toxin synthesis. The insecticidal to (molecular weight, 68,000) from both strains could be prepared by cleaving the protoxin (molecular weight, 135,000) with trypsin, followed by ion-exchange chromatography. The procedure described gave quantitative recovery of toxic activity, and approximately half of the total protein was recovered. Calculations show that these results correspond to stoichiometric conversion of protoxin to insecticidal toxin. The toxicities of whole crystals, soluble crystal protein, and purified toxin from both strains were comparable.
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0004740233 BIOSIS NO.: 198580049128  
DELINEATION OF A TOXIN -ENCODING SEGMENT OF A BACILLUS- THURINGIENSIS CRYSTAL PROTEIN GENE



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JOURNAL: Journal of Biological Chemistry 260 (10): p6273-6280 1985 ISSN: 0021-9258 DOCUMENT TYPE: Article  
RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: Crystals of *B. thuringiensis* ssp. kurstaki HD-1-Dipel contain a MW 134,000 protoxin which can be cleaved by proteolysis to a peptide of MW approx. 70,000; this peptide is lethal to lepidopteran larvae. One analyzed the peptides produced by recombinant *Escherichia coli* strains bearing deletions and fusions of the protoxin gene in order to delineate the portion of the gene which encodes the toxic peptide. The recombinant strains produced the toxic peptide and larger peptides whose size was related to the length of the deleted gene. The amino-terminal 55% of the protoxin protein is sufficient for toxicity. While 2 different gene fusions to the 10th codon allowed the synthesis of toxic polypeptides, fusions to the 50th codon did not. 3' and deletions up to the 645th codon allowed synthesis of the toxic peptide, whereas a deletion to the 603rd codon yielded a non-toxic peptide. Some of the 5' and 3' alterations to the gene caused changes in the proteolytic cleavage patterns of the polypeptides synthesized by *E. coli*, suggesting that the alterations led to conformational changes in the proteins. The presence of different 3' end segments affected the levels of synthesis of the altered crystal proteins.

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0004513711 BIOSIS NO.: 198529042610

BIOSYNTHESIS OF THE INSECTICIDAL TOXIN FROM BACILLUS- THURINGIENSIS -SSP-ISRAELENSIS

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JOURNAL: Abstracts of the Annual Meeting of the American Society for Microbiology 85 p183 1985

CONFERENCE/MEETING: 85TH ANNUAL MEETING OF THE AMERICAN SOCIETY FOR MICROBIOLOGY, LAS VEGAS,

NEV., USA, MAR. 3-7, 1985. ABSTR ANNU MEET AM SOC MICROBIOL. ISSN: 0094-8519 DOCUMENT TYPE: Meeting

RECORD TYPE: Citation LANGUAGE: ENGLISH

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DISSOLUTION AND DEGRADATION OF BACILLUS- THURINGIENSIS DELTA ENDO TOXIN BY GUT JUICE PROTEASE OF THE SILKWORM BOMBYX-MORI

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JOURNAL: Applied and Environmental Microbiology 45 (2): p576-580 1983 ISSN: 0099-2240 DOCUMENT TYPE: Article

RECORD TYPE: Abstract LANGUAGE: ENGLISH

ABSTRACT: The dissolution and degradation of delta-endotoxin (crystal) of *B. thuringiensis* subsp. kurstaki strain HD-1 were investigated. Crystals were dissolved in 0.1 M. phosphate-carbonate-NaOH buffer at pH > 12. Swelling of crystals occurred in the buffer between pH 10 and 11, and crystals dissolved in the same buffer supplemented with gut juice protease of the silkworm *B. mori*. The proteolytic dissolution of crystals occurred after a time lag of several minutes in 0.1 M carbonate-NaOH buffer, pH 10.2. The time lag was not observed when crystals were suspended in the buffer for 30 min. before the addition of protease. After the dissolution of the crystals and further degradation of the solubilized protein, the appearance of a toxic protein with a MW of 59,000, designated P-59, was observed. Lower MW peptides (less than 40,000) showed no toxicity to the silkworm larvae on feeding. Digestion of the 120,000-dalton subunit of the crystal by gut juice protease also produced P-59. These observations suggest the occurrence of a similar process in vivo, i.e., the swelling of crystals due to the alkalinity of gut juice and the production of P-59, dependent on the hydrolysis of swollen crystals by gut juice protease.

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0003883478 BIOSIS NO.: 198375067421

THE MAIN FEATURES OF BACILLUS- THURINGIENSIS DELTA ENDO TOXIN MOLECULAR STRUCTURE

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JOURNAL: Archives of Microbiology 132 (2): p159-162 1982 ISSN: 0302-8933 DOCUMENT TYPE: Article

RECORD TYPE: Abstract LANGUAGE: ENGLISH

ABSTRACT: The crystal-forming proteins (delta-endotoxins) produced by various serotypes of *B. thuringiensis* and toxic for Lepidoptera reveal the same pattern of molecular organization. These proteins (130,000-145,000 MW) contain an N-terminal domain (65,000-85,000 MW) resistant to proteolysis whereas their C-terminal moieties (65,00 MW) undergo an extensive degradation by trypsin that leads to stepwise cleavage off the fragments with MW of 15,000-35,000. The N-terminal domain from serotype V delta-endotoxin is active when introduced into the hemocoel of *Galleria mellonella* larvae. It corresponds to the true toxin normally formed by action of larva proteases on the crystal-forming protein (protoxin). Some differences were found in the properties of the N-terminal domains isolated from the crystal-forming proteins of III, V and IX serotypes, e.g., in their solubility, digestion by subtilisin, MW and the distribution of methionine residues along the polypeptide chains.

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0003025269 BIOSIS NO.: 198070056756

PURIFICATION OF THE INSECTICIDAL TOXIN IN CRYSTALS OF BACILLUS- THURINGIENSIS

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JOURNAL: Journal of General Microbiology 118 (1): p1-12 1980 ISSN: 0022-1287 DOCUMENT TYPE: Article

RECORD TYPE: Abstract LANGUAGE: ENGLISH

ABSTRACT: Crystals were purified from 4 serotypes of the insect pathogen *B. thuringiensis*. Crystals from these serotypes were similar in amino acid and N-terminal analyses, but differed in their toxicity to 2 spp. of Lepidoptera and in their immunological properties. Toxic polypeptides were obtained following trypsin digestion of solutions of the crystals. In 2 strain (serotypes 3 and 9), this fraction contained only 1 polypeptide. Similar results were obtained when dissolved crystals were digested with other proteolytic enzymes or with gut contents from *Pieris brassicae*. The trypsin-resistant polypeptide was further purified by gel and ion-exchange chromatography and had a MW of about 70,000, estimated by gel chromatography a gel electrophoresis. No evidence was obtained for a toxin of lower MW. This purified toxin accounted for most, if not all, of the toxic activity originally present in the crystal solution and was active by injection and ingestion. The purified toxic fraction from serotype 1 appeared to contain 2 polypeptides, one of which corresponded to that found with serotypes 3 and 9. There were no major differences in the composition of crystals from different serotypes of *B. thuringiensis* and it is concluded that trypsin-resistant polypeptide represents the active insecticidal toxin of the crystal.

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0002286900 BIOSIS NO.: 197815004387

SOLUBLE GLYCO PROTEIN INSECT TOXIN FROM THE SPORE COAT OF BACILLUS- THURINGIENSIS

AUTHOR: ARONSON J N; FOX S I

JOURNAL: Federation Proceedings 37 (6): p1824-1978 1978 ISSN: 0014-9446 DOCUMENT TYPE: Article

RECORD TYPE: Citation LANGUAGE: Unspecified

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0002100375 BIOSIS NO.: 197763021231

FURTHER OBSERVATIONS ON THE MODE OF ACTION OF BACILLUS- THURINGIENSIS ON PAPILIO-DEMOLEUS AND SPODOPTERA-LITURA

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ABSTRACT: The influence of reducing substances (oxalic acid, ascorbic acid) and proteolytic enzyme activity of the guts of instar larvae of these insects was investigated to understand better the mode of action of *B. thuringiensis*. The average time required to effect 100% mortality was 1/2 to 1 day in *P. demoleus* as against no mortality in *S. litura*. The highly susceptible *P. demoleus* contained the highest amount of 325 .mu.g/g ascorbic acid as against 194 .mu.g/g in *S. litura*. *S. litura* contained 481.0 .mu.g/g of total phenol and 379 .mu.g/g of OD [orthodihydroxy] phenol and was resistant to *B. thuringiensis* infection. Larvae of *P. demoleus*, highly susceptible to the pathogen, showed noticeably lower amounts of total phenol (215 .mu.g/g) and OD phenol (111 .mu.g/g). *P. demoleus* showed a higher amount of proteolytic enzyme activity (0.425 units) than *S. litura* (0. units). The gut pH of *P. demoleus* was 9.7-10.0 and that of *S. litura* 8.2-8.5. The solubility of the toxic crystal in vivo as a result of the high gut pH in *P. demoleus* probably accounted for the high susceptibility of this species. The presence of large amount of reducing substances like ascorbic acid and phenols favored the S-S bond cleavage. When the S-S bond was broken, the protein moiety of the toxin was acted upon by the proteases. A low pH and low ascorbic acid and phenol contents along with comparatively less proteolytic activity of the gut were the possible factors governing non-susceptibility of *S. litura* to *B. thuringiensis*.

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- 8/7/67 DIALOG(R)File 5:Biois Previews(R) (c) 2004 BIOSIS. All its. reserv.  
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PURIFICATION AND CHARACTERIZATION OF THE ACTIVE FRAGMENT FROM *BACILLUS-THURINGIENSIS* DELTA TOXIN  
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AUTHOR ADDRESS: DEP OF PROTEIN BIOCHEMISTRY, ROCHE RES CENT, HOFFMANN-LA ROCHE INC, NUTLEY, N 07110, USA\*\*USA  
JOURNAL: Biochemical and Biophysical Research Communications 141 (1): p 106-111 1986 ISSN: 0006-291X  
DOCUMENT TYPE: Article RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: Limited tryptic hydrolysis of a partially purified delta-toxin (M<sub>r</sub>=100,000) from *Bacillus thuringiensis* has produced a polypeptide fragment of M<sub>r</sub>=60,000 containing the full biological activity. The fragment was the only polypeptide observed the polyacrylamide-gel electrophoresis of the delta-toxin after treatment with trypsin and could be purified by DEAE-cellulose chromatography. Amino acid and partial sequence analyses indicate that the 60,000 Mr fragment has been derived from the mid-section of the holotoxin peptide; over 80% of Lys, 65% of Pro and 50% of His residues in the holotoxin have been lost in active fragment. This section must contain the active site since its specific insecticidal activity is approximately twice that of t holotoxin. The active fragment shows complete cross-reactivity with the antiserum raised against the native toxin, and appeared to possess higher thermal stability than the mother protein. It provides a powerful tool for studies of the structure involved in the insecticidal activity.
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MODE OF ACTION OF BIPYRAMIDAL DELTA ENDOTOXIN OF *BACILLUS-THURINGIENSIS* -SSP-KURSTAKI HD-1  
AUTHOR: TOJO A (Reprint)  
AUTHOR ADDRESS: INST BIOLOGICAL CONTROL, FAC AGRICULTURE, KYUSHU UNIV, FUKUOKA 812, JAPAN\*\*JAPA  
JOURNAL: Applied and Environmental Microbiology 51 (3): p630-633 1986 ISSN: 0099-2240 DOCUMENT TYPE: Article  
RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: The mode of action of toxic fragment (P-59) derived from bipyramidal-shaped .delta.-endotoxin of *Bacillus thuringiensis* subsp. kurstaki HD-1 on the silkworm *Bombyx mori* was investigated. An enzyme-linked immunosorbent assay showed that there was no translocation of P-59 from the gut lumen to the hemocoel. When membrane vesicles prepared from silkworm midgut were incubated with P-59, normally smooth surface of vesicles became rough, and patch formation was observed on the surface. Vesicles treated with P-59 tended to agglutinate. The vesicle-denaturing activity of a 130,000-dalton subunit protein of bipyramidal toxin was enhanced by treatment with a gut juice protease of the silkworm. P-59 did not cause any uncoupling effect on mitochondria of the silkworm midgut. These results suggest that the attacking site of this toxin is in the mitochondrion but the cell membrane of the susceptible cell.
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0004640586 BIOSIS NO.: 198579059485  
ISOLATION AND ASSAY OF THE TOXIC COMPONENT FROM THE CRYSTALS OF *BACILLUS-THURINGIENSIS* -VAR-ISRAELENSIS  
AUTHOR: DAVIDSON E W (Reprint); YAMAMOTO T  
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JOURNAL: Current Microbiology 11 (3): p171-174 1984 ISSN: 0343-8651 DOCUMENT TYPE: Article  
RECORD TYPE: Abstract LANGUAGE: ENGLISH  
ABSTRACT: The 25-Kdal [kilodalton] fragment of the 28-Kdal toxic protein extracted from *B. thuringiensis* var. *israelensis* crystals is responsible for the insecticidal, cytolytic, hemolytic and mouse-lethal activities of the crude toxin extract. This activity has no relation to the hemolysis produced by other strains of *B. thuringiensis*. This protein was rich in the amino acid Asp and Glu, but did not contain Cys.



126/174	00152900	**Image available** NUCLEOTIDE SEQUENCES CODING FOR POLYPEPTIDES EXERCISING A LARVICIDAL EFFECT IN LEPIDOPTERA SEQUENCES DE NUCLEOTIDIQUES CODANT POUR DES POLYPEPTIDES DOTES D'UNE ACTIVITE LARVICIDE VIS-A-VIS DE LEPIDOPTERES Publication Language: French Fulltext Availability: Detailed Description Claims Fulltext Word Count: 13893 Publication Year: 1988
126/175	00151972	NOVEL BACILLUS THURINGIENSIS STRAINS, METHOD FOR THEIR ISOLATION AND RELATED INSECTICIDAL COMPOSITIONS NOUVELLES SOUCHES DE BACILLUS THURINGIENSIS ,PROCEDE PERMETTANT LEUR ISOLATION ET COMPOSITIONS INSECTICIDES SY RAPPORTANT Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 7983 Publication Year: 1988
126/176	00149737	HYBRID GENES INCORPORATING A DNA FRAGMENT CONTAINING A GENE CODING FOR AN INSECTICIDAL PROTEIN, PLASMIDS TRANSFORMED CYANOBACTERIA EXPRESSING SUCH PROTEIN AND METHOD FOR USE AS A BIOCONTROL AGENT GENE HYBRIDE A FRAGMENT D'ADN CONTENANT LE GENE DE CODAGE D'UNE PROTEINE INSECTICIDE, PLASMIDES, CYNOBACTERIE TRANSFORMEES D'EXPRESSION DE CETTE PROTEINE ET LEUR PROCEDE D'UTILISATION COMME AGENT BIOCIIDE Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 8217 Publication Year: 1988
126/177	00145146	METHOD FOR PRODUCING A HETEROLOGOUS PROTEIN IN INSECT CELLS PROCEDE DE PRODUCTION DE PROTEINES HETEROLOGUES DANS DES CELLULES D'INSECTES Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 4846 Publication Year: 1988
126/178	00138443	AGRICULTURAL-CHEMICAL-PRODUCING ENDOSYMBIOTIC MICROORGANISMS AND METHOD OF PREPARING AND USING SAME MICRO-ORGANISMES' ENDOSYMBIOTIQUES FABRICANT DES PRODUITS CHIMIQUES AGRICOLES, ET LEUR PROCEDE DE PREPARATION ET D'UTILISATION Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 24446 Publication Year: 1987
126/179	00132093	**Image available** CYSTEINE-DEPLETED MUTAINS OF BIOLOGICALLY ACTIVE HUMAN TUMOR NECROSIS FACTOR PROTEINS MUTEINES DEPOURVUES DE CYSTEINE DE PROTEINES DU FACTEUR NECROTICQUE TUMORAL HUMAIN BIOLOGIQUEMENT ACTIF Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 8989 Publication Year: 1986
126	K/180 00129030	**Image available** BACILLUS THURINGIENSIS CRYSTALLINE PROTEIN GENE TOXIN SEGMENT/SEGMENT DE LA TOXINE DU GENE DE LA PROTEINE CRISTALLINE DU BACILLUS THURINGIENSIS Patent: WO 8601536 A1 19860313 Publication Language: English Fulltext Availability: Detailed Description Claims Fulltext Word Count: 11702 Publication Year: 1986
English Abstract: A DNA fragment that codes for the portion of Bacillus thuringiensis crystal protein peptide that is toxic to lepidopteran insects The invention also comprises the DNA... ...invention demonstrates that the disclosed toxin-encoding DNA fragment (referred to herein as the Bacillus thuringiensis crystal protein gene toxin segment) is expressible in recombinant host organisms, and that the "toxin... French Abstract: Un fragment d'ADN code la partie du peptide de proteine cristalline du Bacillus thuringiensis qui est toxique pour des insect lepidopteres. L'invention porte egalement sur les sequences d...la toxine (appelee segment de la toxine du gene de la proteine cristalline du Bacillus thuringiensis peut etre exprimee dans des organismes recombinants hotes, et que le produit de la proteine... Detailed Description: ... this invention relates to an expressible DNA fragment coding for the toxin portion of Bacillus thuringiensis crystal protein. Background of the invention: As is well known Bacillus thuringiensis crystal protein is toxic to the larvae of a number of lepidopteran insects As a result preparations containing Bacillus thuringiensis crystals are used commercially as a highly selective biological insecticidee Unfortunately relatively high manufacturing costs... ...difficult for such insecticides to compete effectively with other commercially available products. Wild-type Bacillus thuringiensis produce crystal protein only during sporulation. Such a growth phase limitation, particularly in an industrial... ...15, 1984 and August 21, 1984, respectively to Schneft and Whiteley disclose expression of Bacillus thuringiensis crystal protein by use o novel recombinant plasmids containing expressible heterologous DNA coding for crystal... ...disclose that genetically engineered bacterial host strains transformed by the novel recombinant plasmids express Bacillus thuringiensis crystal protein protoxin polypeptides. Such genetically engineered bacterial host strains express Bacillus thuringiensis crystal protein protoxin polypeptide at all stages of growth.		



It is now known that in the *Bacillus thuringiensis* subspecies that synthesize lepidopteran toxins the crystal protein crystal is composed of one or more...

...To that end it would be useful to identify the specific segment of a *Bacillus thuringiensis* crystal protein gene that codes for the toxin fragments. It would also be useful to...gene, including some 51 and 31' flanking sequences. The start sites of transcription in *Bacillus thuringiensis* (Bt and Btl) and in *Escherichia coli* (Ec) are indicated as well as the entire...

...It is an object of the present invention to identify the segment of a *Bacillus thuringiensis* crystal protein gene referred to herein as the 'toxin-encoding' segment, which codes for the...

...present invention to determine the DNA sequence of the 'toxin-encoding' segment of a *Bacillus thuringiensis* crystal protein gene. It is a further object of the present invention to demonstrate that a *Bacillus thuringiensis* crystal protein 'toxin-encoding' gene segment is expressible in transformed recombinant host organisms. It is...

...protein gene; FIGURE 2 (Views A and B) shows the DNA sequence of a *Bacillus thuringiensis* crystal protein gene; FIGURE 3 (Views A and B) shows restriction maps of pES1 and...

...shows the construction strategy for the 31-end deletions of a crystal protein @5 *Bacillus thuringiensis* crystal protein. Nucleotides 4140 through 4185 comprise the transcriptional terminator for this gene. FIGURE 3A...that is itself toxic to lepidopteran insects. The phrase 'amino terminal 55%' of the *Bacillus thuringiensis* crystal protein gene' means the amino terminal 645 codons of the crystal protein gene as...

...used herein the phrase 'final 74 codons' means the final 74 codons of the *Bacillus thuringiensis* crystal protein gene as shown in FIGURES 2A and 2B. More specifically the final 74...

...1982; Schnepf and Whiteley, 1981) we have cloned a crystal protein gene from *Bacillus thuringiensis* subspecies *kurstaki* HD-10 (Dipel and have shown that the gene is located on a...well with molecular weights determined for the protoxins from *Bacillus thuringiensis* subsp. *kurstaki* and subsp. *thuringiensis*. In addition the deduced amino acid composition is very similar to the chemically determined amino...

...purified protoxin of *Bacillus thuringiensis* subsp. *kurstaki* (5ga Table I supra.) Knowing that the *Bacillus thuringiensis* crystal protein protoxin can be cleaved to yield a smaller toxin fragment we created recombinant *Escherichia coli* strains bearing deletions and fusions of the crystal protein gene and then analyzed the proteins they produced to delineate the portion of the gene which encodes the toxin peptide. Construction of these 'deletion and fusion' plasmids is discussed in the Materials and Methods section? z112ra, also zge

FIGURES 3 The truncated peptides produced by recombinant hosts transformed by these plasmids are discussed in Examples II through V. Very generally the truncated peptides produced by host strains transformed by the 'deletion' and 'fusion' plasmids indicate that the amino terminal 55% of the crystal protein gene encodes sufficient information to produce a lepidopteran toxin. More specifically the 'deletion' and 'fusion' results indicate that deletions to the 50th codon from the 51' end of the gene or to the 603rd codon from the 31' end abolish toxicity while deletions to the 10th codon from the 51' end, or to the 645th codon from the...sequence was required to determine which portion of the crystal protein gene coded for the 'toxin' portion of the crystal protein. The DNA sequence for the remainder of the gene was...

...from bases 7845 to 3831 in FIGURE 1B was sequenced primarily by the DNase I deletion method of Hong (1982) (z1je numbered sites in FIGURE 1B) but some gaps in the coding strand sequence were filled in by obtaining deletions through partial *Sau3A*I digestion (asterisks in FIGURE 1B). The complete sequence was determined for both...protoxins from *Bacillus thuringiensis* subsp. *kurstaki* (Bulla, et al., 1981) and subsp. *thuringiensis* (Hubert et al., 1981). The deduced amino acid composition is very similar to the...the crystal protein gene encodes a toxic peptide and that the 31' end of the toxin-encoding portion was in the *HindIII*-E fragment. To determine the 31' end of this region more precisely a number of deleted plasmids were constructed. See FIGURE 6A. The crystal

protein encoding sequence in these plasmids terminated... Claim... and 2B. 2a A composition of matter comprising the amino terminal 55% of a *Bacillus thuringiensis* crystal protein gene as shown in FIGURES 2A and 2B.

39 A DNA sequence having...

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METHODS AND VECTORS FOR TRANSFORMATION OF PLANT CELLS PROCEDES ET VECTEURS POUR LA TRANSFORMATION DE CELLULES VEGETALES

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